

Influence of The Experimental Design on Shake Flask Culture of Acidithiobacillus (At.) ferrooxidans Using Sulfur as Substrate

Samir Daniel, Anne-Gwénaëlle Guezennec, Agathe Hubau, Douglas O.

Pino-Herrera, Eric Olmos

▶ To cite this version:

Samir Daniel, Anne-Gwénaëlle Guezennec, Agathe Hubau, Douglas O. Pino-Herrera, Eric Olmos. Influence of The Experimental Design on Shake Flask Culture of Acidithiobacillus (At.) ferrooxidans Using Sulfur as Substrate. 63rd Annual Conference of Metallurgists, Aug 2024, Halifax, Canada. hal-04533904

HAL Id: hal-04533904 https://brgm.hal.science/hal-04533904v1

Submitted on 5 Apr 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

IBS 2024 Abstract

Influence Of The Experimental Design On Shake Flask Culture Of Acidithiobacillus (At.) ferrooxidans Using Sulfur As Substrate

Samir Daniel¹, Anne-Gwénaëlle Guezennec¹, Agathe Hubau¹, Douglas Pino-Herrera¹, Eric Olmos²

¹BRGM, F-45060 Orléans, France

²LRGP, UMR CNRS 7274, F-54000 Nancy, France.

In bioleaching field, shake flask is a common experimental device that has been used in numerous research works to cultivate microorganisms. However, none of these studies has considered if the agitation and aeration conditions applied to these cultures were sufficient to avoid O₂ transfer limitations and to promote efficient solid particles dispersion, two key mechanisms in bioleaching experiments. Our study aimed at assessing the influence of the design of shake flask experiments on the growth of At. ferrooxidans using elemental sulfur (S₀) as substrate. Cultures of At. ferrooxidans in 0Km medium with 1% sulfur were incubated at 30°C for 10 days in duplicates. Different culture volumes (V, 75 to 150 mL), agitation rates (N, 101 to 350 rpm) and orbit diameters (D_0 , 19 to 25 mm) were used to vary the gas-liquid mass transfer coefficient ($k_I a$, from 8 to 41 h⁻¹) and the power dissipation per unit of volume $(P_0/V, \text{ from 21 to 982 W/m}^3)$ from one culture to another. Daily monitoring of the cultures included the measurement of pH, dissolved oxygen concentration and biomass (concentration of bacteria in the liquid phase). Dissolved sulfate concentration was also measured by IEC at the beginning and the end of the culture. At. ferrooxidans growth (characterized by μ_{max}) was improved with the increase of $k_L a$ and P_0/V while S⁰-biooxidation rate was increased. Moreover, whatever the experimental conditions, dissolved oxygen concentration was always above 85% of saturation, indicating that there was no limitation of the gas-liquid mass transfer in the range of $k_L a$ used for the study. Increasing P_0/V for a given value of $k_L a$ improved the growth rate while it remained the same for a fixed P_0/V and different $k_L a$. As an example, for a $k_L a$ equal to 16 h⁻¹, μ_{max} increased from 0.028 to 0.035 h⁻¹ when P_0/V increased from 34 to 198 W/m³. On the contrary, for P_0/V equal to 200 W/m³, μ_{max} remained close to 0.035 h⁻¹ when k_{La} was increased from 16 to 23 h⁻¹. It was also observed that μ_{max} reached a plateau when P_0/V was above 600 W/m³. Interestingly, at the plateau, the agitation rate was higher than the theoretical critical rate N_c required to achieve a complete suspension of the sulfur particles in the liquid, which was not the case for the other cultures. These results show that microbial growth of At. ferrooxidans on S^0 was mainly influenced by particles dispersion and

not by gas-liquid mass transfer. In this case, P_0/V and N_c are two key factors that must be considered in the design of the experiments. The study confirms that operating conditions such as *N*, *Do* and *V*, which influence the aeration and homogenization of the culture in shake flask, must be carefully selected to avoid limiting bacterial growth.