



HAL
open science

Transfer and degradation of polyacrylamide based flocculants in hydrosystems: a review

Anne-Gwenaëlle Guezennec, Caroline Michel, Kathy Bru, Solène Touzé, Nicolas Desroche, Inès Mnif, Mikael Motelica-Heino

► To cite this version:

Anne-Gwenaëlle Guezennec, Caroline Michel, Kathy Bru, Solène Touzé, Nicolas Desroche, et al.. Transfer and degradation of polyacrylamide based flocculants in hydrosystems: a review. Environmental Science and Pollution Research, 2015, 22 (9), pp.6390-6406. 10.1007/s11356-014-3556-6 . hal-01071342

HAL Id: hal-01071342

<https://brgm.hal.science/hal-01071342>

Submitted on 6 Oct 2014

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **Title: Transfer and degradation of polyacrylamide based flocculants in hydrosystems: a review**

2

3 **Authors:** A. G. Guezennec¹, C. Michel¹, K. Bru¹, S. Touze¹, N. Desroche², I. Mnif³, M. Motelica-Heino⁴

4 ¹ BRGM

5 ² Nexidia SAS, 15 Rue de Mayence 21000 DIJON France

6 ³ Université de Nice

7 ⁴ ISTO, UMR 7327 CNRS-Université d'Orléans, 1A rue de la Férollerie, 45071 Orléans cedex 2

8

9 **Corresponding author:**

10 Anne-Gwénaëlle Guezennec – a.guezennec@brgm.fr

11

12 **Abstract**

13 The aim of this review was to summarize information and scientific data from the literature dedicated
14 to the fate of PAM-based flocculants in hydrosystems. Flocculants, usually composed of
15 polyacrylamides (PAMs), are widely used in several industrial fields, particularly in minerals extraction,
16 to enhance solid/liquid separation in water containing suspended matter. These polymers can contain
17 residual monomer of acrylamide (AMD) which is known to be a toxic compound. This review focuses
18 on the mechanisms of transfer and degradation, which can affect both PAM and residual AMD, with a
19 special attention given to the potential release of AMD during PAM degradation. Due to PAM ability to
20 adsorb onto mineral particles, its transport in surface water, groundwater and soils is rather limited and
21 restricted to specific conditions. PAM can be also subject of biodegradation, photodegradation and
22 mechanical degradation but most of the studies report slow degradation rates without AMD release. In
23 the contrary, the adsorption of acrylamide onto particles is very low, which could favor its transfer in
24 surface waters and ground water. However AMD transfer is likely to be limited by quick microbial
25 degradation.

26

27 **Key words:** flocculation, polyacrylamide, acrylamide, environmental hazards, transfer, adsorption,
28 photodegradation, biodegradation, amidase

29

30 1. Introduction

31 Flocculants are widely used in several industrial fields (mineral extraction, chemical industry, food
32 processing industry, treatment of drinking water, etc.) to enhance solid/liquid separation in water
33 containing suspended matter (Heath *et al.*, 2006, Harford *et al.*, 2011). In most cases, flocculants are
34 polymers composed of polyacrylamide (PAM) (Taylor *et al.*, 2002). PAM is a polymer with high
35 molecular weight, synthesized from acrylamide (AMD) and acrylic acid/acrylate (AA) (Scott *et al.*, 1996;
36 Caulfield *et al.*, 2002). The use of flocculants enables to increase the recycling rate of process water
37 and to decrease sludge volumes and thus the surface of settling ponds. Those ponds are open
38 systems with possible exchanges into the near aquatic environment (water percolation towards soils
39 and aquifers, water seepages in natural streams or rivers...), which can lead to the dissemination of
40 flocculants into surface and groundwaters (de Rosemond & Liber, 2004; Liber *et al.*, 2005). The
41 question of flocculants safety is now arising as a new key-environmental issue. The potential
42 environmental hazard does not concern the polyacrylamide itself, which is generally considered as not
43 toxic (Andersen, 2005), but the products of PAM degradation and the residual acrylamide contained in
44 the flocculants due to incomplete polymerization process (Labahn *et al.*, 2010; Young *et al.*, 2007;
45 Caulfield *et al.*, 2002). AMD is reported to be a neurotoxin to humans (McCollister *et al.*, 1964). AMD is
46 also classified as a carcinogenic (level 2), mutagenic (level 2) and reprotoxic (level 3) compound
47 (World Health Organization 1985; Molak, 1991). This review focuses on the fate of PAM-based
48 flocculants in hydrosystems, with a major emphasis on the mechanisms of transfer and degradation
49 which can affect both PAM and associated AMD. The section dedicated to transfer phenomenon will
50 especially examine transport and reactivity with solid matrices. In the section devoted to degradation,
51 abiotic and biotic factors will be discussed with a specific emphasis on potential hazardous AMD
52 release during PAM degradation.

53

54 2. Polyacrylamide and acrylamide in flocculation processes and potential environmental 55 hazards

56 The word polyacrylamide and the acronym "PAM" (for polyacrylamide) refer to a group of water
57 soluble molecules formed by polymerization of acrylamide monomers. PAMs have been used for
58 decades in a variety of industries and have proven particularly effective in facilitating the solid-liquid
59 separation for waste and drinking water treatment as well as for the clarification of various types of

60 effluents. High molecular weight PAMs (10^6 to 10^7 g.mol⁻¹) work as flocculants by causing suspended
61 particles to bind together to form larger aggregates which can settle out quickly from solution leaving a
62 clear supernatant (Heath *et al.*, 2006). By increasing solid-liquid separation, PAMs improve water
63 clarification processes and water quality prior to discharge, re-use or disposal. In France, for instance,
64 the consumption of PAM-based flocculants is about 15 kt per year. In particular, the aggregate
65 production uses about 900 t/y of PAM-based flocculants (i.e. 6% of the total consumption) as
66 clarification aid (UNPG 2013).

67 High molecular weight PAMs can be modified to develop non-ionic, anionic, or cationic properties for
68 specific uses (Scott *et al.*, 1996). However for environmental applications which are described in this
69 article, anionic PAMs are preferred because of their extremely low aquatic toxicity compared to
70 cationic forms (Hamilton *et al.*, 1994; de Rosemond & Liber, 2004; Liber *et al.*, 2005). Common anionic
71 PAMs are usually produced by copolymerization of acrylamide and acrylic acid or one of its salts (see
72 Fig.1). Their molecular weights vary between $3 \cdot 10^6$ and $2 \cdot 10^7$ Da. The most polyvalent anionic PAM
73 contains around 30% of acrylate (in mol).

74 Anionic PAMs used in flocculation applications are water-soluble and have little if any cross-linking.
75 The molecules, when dissolved in water, are nominally linear, although they may be coiled or curled to
76 varying degree due to either substitution along the chain or as a result of electrolytes in the solvating
77 water (Sokja *et al.* 2007).

78 PAMs can be manufactured to have different molecular weights and charge densities by varying the
79 reaction parameters and/or the relative quantities of reagents used. The charge density of PAM is
80 often expressed as percent anionic, referring to the percentage of its monomers that contain a
81 charged functional group. The variations can result in significant differences in the extent to which
82 PAM will bind to different types of particles, and thus influence the flocculation efficiency. Even among
83 products that contain anionic PAM as their active ingredient, performance can vary substantially if the
84 PAM charge densities or molecular weights are different.

85 One of the primary areas of concern associated with the use of PAMs is the presence of residual
86 monomer of AMD in the polymer product, due to incomplete polymerization process. AMD is solid at
87 room temperature, highly soluble in water, has a low potential to partition to organic matter, and has a
88 low volatilization potential in water. Acrylamide, often referred by the acronym AMD, is considered to
89 be a likely human carcinogen and neurotoxin (EPA US 2010). All PAMs contain some level of residual

90 AMD. However, the amount of residual AMD can vary substantially depending on what measures were
91 taken during the manufacturing process to maximize the extent of polymerization. Within the EU, all
92 PAMs are required to contain less than 0.1% (w/w) of residual AMD (European Parliament 1999)
93 unless they are classified and labeled as a Category 2 carcinogen (European Parliament 2006). More
94 stringent thresholds are also set depending on additional regulations covering specific uses like, for
95 example, when PAM-based flocculants are used for the treatment of drinking water (no more than
96 0.05% of residual AMD). Most of the commercial PAMs products complied with this requirement, even
97 when they are not marketed for drinking water treatment. Knowledge about the fate of PAM and
98 associated residual AMD in hydrosystems (transfer, reactivity...) is limited or non-existent in the field
99 of the use of PAM as a flocculating agent for the treatment of waters. This monomer is water soluble
100 and unlikely to adsorb to organic and inorganic soil components, potential for leaching and
101 groundwater contamination are often considered as the major environmental risk associated with the
102 use of PAMs as flocculants. Several publications have discussed the aquatic toxicity of anionic PAM in
103 the framework of the struggle against agricultural soil erosion where PAMs are used for soil
104 stabilization (Barvenik 1994; Seybold 1994; Entry *et al.* 2002; Sojka *et al.* 2000, 2007). These articles
105 have disregarded its toxicity potential, based largely on theoretical grounds such as the expectation
106 that toxicity would be mitigated by the complexation with dissolved organic matter (Goodrich *et al.*
107 1991), because any PAM leaving the field would be rapidly adsorbed to soils (Lentz *et al.* 2002), and
108 also because the size of the molecule would prevent passage across biological membranes (Stephens
109 1991). Few authors have also studied the fate of associated residual AMD when PAMs are used for
110 soil stabilization and demonstrated that in this context AMD undergoes a quick degradation, which
111 prevents its dispersion in the environment (Labahn *et al.*, 2010; Arrowood, 2008, Shanker *et al.*, 1998;
112 Lande *et al.*, 1979). Even if these studies are an important source of information, their results and
113 conclusions cannot be transposed directly to the field of PAM-based flocculants because the
114 phenomenon that can affect the fate of PAM and associated AMD are slightly different and because
115 PAMs used for both applications have different properties (molecular weight and charge densities). It
116 must also be noted that in the field of flocculation, the main vector of dispersion of PAM and AMD in
117 the environment is water, whether surface or groundwater. In the following parts of this article, the
118 mechanisms of PAM and AMD transfer and degradation will be thoroughly reviewed and their potential
119 impacts on the fate of PAM-based flocculants in hydrosystems will be assessed.

120

121 **3. Transfer of PAM –based flocculants in hydrosystems**

122 **3.1. Sorption mechanisms**

123 A variety of PAM and AMD molecular and environmental properties interact to affect their transfer in
124 hydrosystems. These interactions can be significant given the range of PAM use in the environment
125 and the number of environmental and application factors that can vary. Studies related to the
126 environmental fate of PAM have been conducted in the framework of several uses of PAMs like soil
127 stabilization and infiltration enhancement in agriculture (Entry *et al.* 2008; Spofford and Pfeiffer 1996).
128 Advances in theoretical chemistry have contributed to improvements in PAM performances through
129 design of molecular conformations optimally suited to given industrial and environmental applications
130 (Bicerano 1994; Bourani 1998; Bouranis *et al.* 1995; Chamberlain and Cole 1996). Nevertheless,
131 many of the general principles from these studies are worth noting as a framework for understanding
132 the environmental behavior of anionic high molecular weight PAMs. Additionally a good deal of
133 information on the effects of coagulants and flocculants comes from the wastewater treatment
134 literature, although often from polymers and compounds other than PAM, or in the case of PAM, often
135 from cationic or nonionic formulations.

136

137 ***Mechanisms of PAM-based flocculants adsorption on mineral particles***

138 The adsorption of PAMs from aqueous media onto mineral surfaces has been largely described in the
139 literature. For instance the adsorption of PAM on soil and clay mineral surfaces has been
140 demonstrated to be rapid and irreversible in several studies although the degree of adsorption itself is
141 dependent on PAM conformation, soil or mineral properties, and soil solutions characteristics
142 (Hollander *et al.* 1981; Nabzar *et al.* 1984; Nabzar *et al.* 1984; Nabzar *et al.* 1988; Nadler *et al.* 1992;
143 Pradip and Fuerstna 1980; Tanaka *et al.* 1990; Stutzmann and Siffert 1977; Theng 1982; Tekin *et al.*
144 2005, Tekin *et al.* 2006; Deng *et al.* 2006; Tekin *et al.* 2010). As PAM adsorption implies multi-
145 segment interactions, there is a small probability that all train segments are simultaneously detached
146 from the surface (Theng 1982). Once adsorbed, less than 3% of the adsorbed PAM is removed from
147 clay minerals after four consecutive washes (Deng *et al.* 2006). Moreover according to Nadler *et al.*
148 (1992), after adsorption and drying, little desorption of anionic PAM from soil occurs. Michaels and
149 Morelos (1955) reported that 20% hydrolysis of PAM provided the greatest degree of chain extension,

150 facilitating its adsorption. As PAMs are drawn to mineral particle surfaces, surface-adsorbed water is
151 driven away because of the stronger attraction for the polymer (Parfitt and Greenland 1970). Sorption
152 isotherms of PAM generally fit with the linear form of Langmuir equation in comparison with the
153 Freundlich isotherm (Lu *et al.* 2002; Deng *et al.* 2006; Tekin *et al.* 2005; Tekin *et al.* 2006; Tekin *et al.*
154 2010). Additionally results of sorption experiments and parameters of Langmuir isotherm found in early
155 studies are consistent with a favorable adsorption and a high affinity of PAM to soil minerals. This high
156 affinity is likely due to the mechanisms of multi-segment sorption of the long chain of PAM (Theng
157 1982).

158 Stutzmann and Siffert (1977) showed that the fixation mechanism of partly hydrolyzed PAM onto clays
159 in a weakly acidic media is a chemisorption phenomenon. The anionic polyelectrolyte is in fact sorbed
160 exclusively on the external surface of the clay, where organic molecules undergo protonation and are
161 retained by ionic interaction. Charge transfer apparently depends on the polarizing power of the
162 exchangeable cation of the clay and a linear relationship was found between the polarizing power of
163 the alkali elements and the quantity of material sorbed.

164 Hydrogen bonds are thought as the main mechanism of adsorption of polymers on mineral surfaces.
165 Adsorption studies of clay-organic complexes have shown that two kinds of H-bonding are possible.
166 One type of H-bonding can occur between the amide groups of PAMs and the protonated aluminol
167 groups on the edge of clays (Laird 1997). Depending on pH value, amide group of the PAM can
168 adsorb either on silanol anchoring sites (at pH 4) or on aluminol anchoring sites (at pH 10) (Pefferkor
169 1987). Another type of H-bonding was proposed between carbonyl groups and edge aluminols (Laird
170 1997). Infrared analysis of clay-organic complexes has shown H-bonding between amide groups of
171 PAM and water molecules in the hydration shell of exchangeable cations of a smectite (Deng *et al.*
172 2006). Finally, ion-dipole interaction has been proposed in controlling the adsorption of PAM on
173 smectite. This interaction takes place between exchangeable cations of smectite and the carbonyl
174 oxygens of amide groups (CONH₂) of PAM (Deng *et al.* 2006).

175 For anionic PAMs, adsorption on kaolinite, illite and quartz is enhanced in the presence of divalent
176 cations through a cationic bridge process. Cationic bridges take place between divalent cations and
177 negative groups of anionic polymer on one hand and divalent cations and negatives sites of mineral
178 surfaces on the other hand. This binding mode occurs at high concentrations of divalent cations (Ca²⁺,

179 Mg^{2+}), and enhances the sorption of anionic polymers on negative mineral surfaces (Theng 1982
180 Chiappa *et al.* 1999).

181 Adsorption of PAMs on the mineral fraction is correlated with size of particles. Clays being the mineral
182 components with the lowest size have more affinity for PAMs than larger particles like sand and silt (Lu
183 *et al.* 2002; Gravelieng *et al.* 1997; Malik and Letey 1991). Clays with a high specific outer-surface
184 area such as montmorillonite and smectite are better PAMs adsorbents than clays with low specific
185 outer-surface areas such as kaolinite and illite (Lu *et al.* 2002; Deng *et al.* 2006; Inyang and Bae
186 2005). Additionally the charge repulsions confer to anionic PAM coil an extended conformation making
187 difficult its entry in the interlayer space of clay minerals (Stuzmann and Siffert 1977). Nevertheless, a
188 small amount of PAM molecules has been shown to penetrate in the interlayer space of
189 montmorillonite due to its large interlayer space (Inyang and Bae 2005). It was also shown that
190 adsorption of PAM increased on montmorillonite when the degree of hydrolysis of PAM increased.
191 PAM indeed tends to take the conformation of an extended chain when it is highly hydrolyzed.
192 Therefore the adsorption of PAM depends on the configuration and the volume of the solvated
193 macromolecule (Stuzmann and Siffert 1977). In contrast, a change of molecular weight has practically
194 no influence upon the adsorption ratio.

195

196 ***Physico-chemical parameters influencing the adsorption of PAM: pH, ionic strength and*** 197 ***temperature***

198

199 The pH value influences both the structure of adsorbent, adsorbate and adsorption mechanism of
200 PAMs on solid surfaces. The effect of pH on the polymer consists on load changes of functional
201 groups by the phenomenon of protonation or deprotonation. At a high pH value, the carboxyl group (-
202 COOH) of polymer can be dissociated as follows: ($-COOH = -COO^- + H^+$), giving the polymer a
203 negative charge. Furthermore, amides groups can be hydrolyzed and generate carboxyl groups (-
204 $CONH_2 = -COOH + COO^-$) (Deng *et al.* 2006). The net charge of natural solids like clays and silicates
205 also depends on the pH. For instance the edge sites of aluminosilicates minerals are amphoteric
206 groups, able to fix or release protons. Thus, the surface charge can be positive when pH is inferior to
207 PZC (point of zero charge), neutral when pH is equal to PZC, or negative when pH is superior to PZC.
208 pH values inferior to the PZC induce the protonation of aluminol groups which confers to solid surfaces

209 a positive net charge. Inversely, at pH values superior to the PZC, aluminols sites and silanols sites
210 are deprotonated, the surface charge becomes negative. Unlike cationic PAMs, the sorption of anionic
211 PAMs is promoted when the surface is positively charged. This is mainly due to the reduction of
212 electrostatic repulsions between negatives charges of polymers and the negative charges of the
213 mineral surface. When $pK_a[\text{COOH}/\text{COO}^-] < \text{pH} < \text{PZC}$, attraction of carboxylic groups towards
214 aluminol groups is enhanced since PAM is negatively charged and the surface is positively charged.
215 When $\text{pH} > pK_a[\text{COOH}/\text{COO}^-]$ and $\text{pH} > \text{PZC}$, the complete dissociation of carboxylic groups
216 reinforces the negative charges of PAM and the surface is negatively charged. This results in the
217 decrease of the adsorption of PAM (Lee *et al.* 1991). The adsorption of PAM tends toward zero when
218 the pH value still increases because of the very large electrostatic repulsions and the sharp reduction
219 of AlOH sites (Deng *et al.* 2006; Lee *et al.* 1991).

220

221 The ionic strength is also an important parameter. Benhur *et al.* (1992) found that the adsorption of
222 PAMs onto illite and montmorillonite clays was generally in the order cationic > nonionic > anionic
223 regardless of the electrolyte content of the solvating water. Water that was more saline and sodic
224 reduced the adsorption of cationic and nonionic PAMs but increased adsorption of anionic PAMs. The
225 results obtained in saline media may be explained by a size decrease of the PAM macromolecules,
226 allowing a closer approach to the surface of the mineral substrate, and by a chain lengthening as the
227 degree of hydrolysis increases, induced by the electrostatic repulsions between the COO^- groups of
228 the medium. Aly and Letey (1988) found that the adsorption of anionic PAMs in water of electrical
229 conductivity (EC) 0.7 dS m^{-1} was greater than for water of EC of 0.05 dS m^{-1} . Entropy change as water
230 is displaced is an important actuating force in bringing about adsorption of negative and nonionic
231 polymers to negatively charged clay surfaces (Lyklema and Fleer 1987; Theng 1982). As the
232 adsorption of anionic PAMs on mineral surfaces is governed by the competition between polymer
233 attractive interactions with solid surfaces and repulsive electrostatic forces (Lecourtier *et al.* 1990), the
234 sorption of anionic polyacrylamide is increased when electrostatic repulsions are screened by cation
235 bonding on the negatively charged surface sites (Lu *et al.* 2002; Lu and Wu 2003). It has also been
236 demonstrated that the adsorption of anionic PAM on the edge surfaces of kaolinite is strongly
237 dependent on salinity (Lee *et al.* 1991). Indeed, two effects have been reported. First, the adsorption
238 of mono and divalent cations from the electrolyte on the negatively charged surface screens the

239 electrostatic repulsion between anionic PAM and the mineral surface. Secondary, since divalent
240 cations are involved in the cationic bridge the sorption saturation of PAM is higher in the presence of
241 divalent cations than in the presence of monovalent cations (Lu *et al.* 2002). Moreover cation
242 enhancement of sorption is more effective in fine-textured soils than in coarse-textured soils.

243

244 Finally, studies conducted on the effect of temperature on the adsorption of anionic PAM have shown
245 contradictory results. Increasing temperature (from 20°C to 60°C) was found to increase the
246 adsorption of anionic polyacrylamide on kaolinite, sepiolite and perlite (Mpofu *et al.* 2004; Tekin *et al.*
247 2005; Tekin *et al.* 2006; Tekin *et al.* 2010). This effect was attributed to two phenomena. Firstly,
248 increasing temperature results in a break of polymer-water hydrogen bonds that promotes the
249 polymer-polymer interactions. The polymer thus adopts a more compact conformation that will adsorb
250 more than the expanded one (Mpofu *et al.* 2004). Secondly, increasing temperature may produce a
251 swelling effect within the internal structure of the clay so the PAM can penetrate further (Tekin *et al.*
252 2005).

253

254 **3.2. PAM Transfer**

255 ***Transfer in water***

256 Both molecular properties and product preparation can influence PAMs behavior in hydrosystems
257 (Callebaut *et al.* 1979). Results from monitoring of PAM concentrations in the drainage water of a
258 tunnel in Sweden where PAM was used showed that PAMs leaked out, both in connection with the
259 injection and in connection with after-injection using other grouting agents (Weideborg *et al.* 2001).

260

261 ***Flocculation***

262 The general principles of flocculation involving PAMs can be easily drawn from the abundant literature
263 dedicated to the adsorption of PAMs from aqueous media onto natural mineral surfaces (Greenlan
264 1963; Bottero and Lartiges 1992; Barvenik 1994; Theng 1979; Theng 1982; Sokja *et al.* 2007; Benhur
265 *et al.* 1992; Haschke *et al.* 2002; Lu *et al.* 2002; Lurie and Rebhun 1997). In an aqueous solution with
266 sufficient electrolytes, coulombic and van der Waals forces attract suspended particles to anionic
267 PAM. These surface attractions stabilize the structure of the aggregates by enhancing particle
268 cohesion. PAM efficiency as flocculants is not only due to adsorption phenomena. As can be seen on

269 Fig. 2, adsorbed PAM can present three different conformations: “train” conformation, “loop”
270 conformation or “tail” conformation. Loops and tails can adsorb onto other particles enabling gathering
271 of several particles in a process known as “polymer bridging”. This phenomenon increases with the
272 molecular weight of PAMs.

273 The charge density plays also an important role in flocculation process. In anionic PAMs, carboxylic
274 groups are distributed regularly on the molecular chain, which enhances the polymer extension by
275 electrostatic repulsion. On another hand, those charges decrease the adsorption onto natural particles
276 charged negatively (like clay particles). The use of PAMs poorly charged and with a high molecular
277 weight usually decreases the competition between both phenomena and favor bridging processes.
278 Small amounts of dissolved Ca^{2+} or Mg^{2+} can also help by shrinking the electrical double layer
279 surrounding particles and bridging the anionic surfaces of particles and anionic PAM molecules. The
280 mechanisms by which flocculation of suspended sediments takes place in the presence of
281 polyelectrolytes was studied by Ruehrwein and Ward (1952) who associated this effect with
282 stabilization of soil and resistance to dispersion. Benhur and Letey (1989) and Benhur *et al.* (1989)
283 attributed this effect to the mechanism that reduced particle dispersion when sprinkler irrigating with
284 PAM, which in turn reduced surface sealing and slowed the reduction of infiltration rate.

285 At temperatures from 6 to 29 °C, flocculation for a variety of inorganic and polymer compounds tended
286 to be slower and flocs tend to be smaller than at higher temperatures (Fitzpatrick *et al.* 2004; Hanson
287 and Cleasby 1990). Furthermore, floc strength seems to vary with the shear conditions of the flow
288 media in which flocs are formed. Larger flocs formed at higher temperatures are more easily disrupted
289 and less capable of reformation than flocs formed at lower temperatures (Fitzpatrick *et al.* 2004;
290 Yeung and Pelton 1996; Yeung *et al.* 1997).

291 PAM solutions usually promote flocculation of suspended solids at low concentrations but can have
292 the opposite effects at higher concentrations, where the large physical domain of the macromolecules
293 themselves interfere with flocculation and actually stabilize dispersed suspensions as viscosity
294 increases. When there is too much of PAM added in the suspension, a compression of the polymer
295 can also be observed on the particle and there is not enough sites available for bridging, which
296 decreases the flocculation efficiency. The concentration, at which the effects reverse, depends on
297 several factors including PAM molecular weight, charge and conformation of the molecule, size and
298 chemistry of the dispersed solids, and chemistry of the water, particularly its electrical conductivity

299 (Sato and Ruch 1980). The treatment of an effluent containing suspended particles requires
300 determining precisely the optimal concentration of PAM which must be added in order to perform an
301 efficient flocculation. This concentration is usually proportional to the particles concentration and is
302 almost always below 1 mg.L^{-1} of suspension.

303

304 ***Transfer with suspended matter***

305 A study measured PAM concentrations in 167-m-long PAM-treated tail ditch that received this runoff
306 (Lentz *et al.* 2002). The soil was a Portneuf silt loam (coarse-silty, mixed, superactive, mesic
307 applications Durinodic Xeric Haplocalcid) with a 1.5% slope. PAM was adsorbed to soil and removed
308 solution as the streams traversed the soil-lined channels. The removal application rate increased with
309 the stream sediment concentration. Additionally a previous study reported that PAM applications of 1
310 to 2 mg.L^{-1} controlled furrow erosion similarly on 1.5% sloping fields (Lentz and Sojka 2000). The
311 linear PAM molecule assumes the form of a hydrated random coil when dissolved in water. Solvated
312 PAM molecules in the furrow stream collide with soil particles when treated water infiltrates into soil or
313 when turbulent flow drives the molecules against entrained sediment or the wetted soil perimeter. The
314 dissolved high molecular weight polymers are in fact readily adsorbed to soil particles via electrostatic,
315 hydrogen, and chemical bonding, and by displacement of inner solvation-sphere water molecules
316 (LaMer and Healy 1963; Mortland 1970; Jin *et al.* 1987; Malik *et al.* 1991; Laird 1997). As a result,
317 incoming PAM is bound to soil in the upper 1 to 5 cm of the profile (Malik *et al.* 1991). Dry soil adsorbs
318 more polymers than wet soils because sorbed water reduces the number of potential soil binding sites
319 (Chang *et al.* 1991). PAM is adsorbed to particulate matter and flocculates soils suspended in water.
320 Polymer adsorption on soil occurs rapidly during the first minutes of exposure but may continue at a
321 reduced rate for several hours or days (Van de Ven 1994). In batch tests (soil, water, and dissolved
322 PAM mixed in a shaker), Nadler *et al.* (1992) reported that little or no polymer desorbed from the soil
323 while it remained wet, and the polymer became irreversibly bonded to the soil upon drying. In flowing
324 systems, Lee and Fuller (1985) found that polymer adsorption rate decreased with increasing velocity
325 of flow. Polymer desorption did not occur under quiescent conditions but was observed when the
326 adsorbent material was subjected to flow shear. Desorption increased with the increasing flow velocity.
327 These authors also concluded that adsorbed PAMs do not penetrate soil aggregates, but only coat
328 and stabilize their surfaces. Malik and Letey (1991) and Nadler and Letey (1989) used tritium-labeled

329 polymers to determine sorption isotherms of several types of polyanions on Arlington sandy loam
330 (coarse-loamy, mixed, thermic Haplic Durixeralfs). They interpreted their results as showing that
331 polymer sorption was restricted to the external surfaces of soil aggregates. A similar conclusion was
332 reached by ElHardy and Abd ElHardy (1989) who saw only limited intrusion of high molecular weight
333 PAMs into soil aggregates. Aly and Letey (1988) showed that the water quality of the solvating water
334 used to apply the polymers also greatly influenced the degree of adsorption.

335 Soils with a high clay or silt content, fine texture and low organic matter content had high sorptive
336 affinity for PAMs. According to Lu *et al.* (2002), organic matter decreased the amount of anionic PAM
337 adsorption on soil. The negative effect of organic matter on PAM sorption was attributed to the
338 reduction of accessible sorption sites by cementing inorganic soil components to form aggregates and
339 to the enhancement of electrostatic repulsion between PAM and soil surface by its negatively charged
340 functional groups. The length of the polymer chain and large number of adsorption sites along the
341 molecule contribute to PAM's adsorption always attached to the adsorption surface, preventing
342 removal of the molecule (Nadler and Letey 1989). Because of the demonstrated high sorptive ability
343 and low mobility (Malik and Letey 1991; Malik *et al.* 1991), PAM is generally regarded as incapable of
344 penetrating soil more than a few centimeters from the soil surface. This, considered in light of the
345 relatively slow sorption kinetics of the large PAM molecule (Lu *et al.* 2002), has led to recognition that
346 the depth of PAM penetration depends on PAM properties, application method, and the soil and water
347 properties present in the application scenario (Lu and Wu 2003). Using anionic PAMs of 10-15 mg
348 mol⁻¹, Lu and Wu (2003) found that PAM penetration depth was about one-eighth to one-half of the
349 water penetration depth, with a particularly high PAM retention in the top few centimeters of the soil.
350 The PAM retained in the top 0-2 mm of soil ranged from 16% to 95% of the total applied. PAM
351 retention was greater at shallow depths when solution contact with the soil was favored by pore
352 arrangement, contact time, and drier soil conditions on addition of PAM solutions.

353

354 **3.3. Transfer of AMD**

355 Mobility of AMD in soils has been little studied. Adsorption of AMD on kaolinite, montmorillonite and
356 anionic, cationic, and hydrophobic resins is insignificant (Brown *et al.* 1980), at acrylamide
357 concentrations between 500 µg L⁻¹ and several mg L⁻¹. Arrowood (2008), who studied the adsorption
358 of acrylamide on different soils (silica sand, coarse sandy soil, loam soil) at AMD concentrations lower

359 than those tested by Brown *et al.* (1980), observed acrylamide adsorption on the three soils with better
360 affinity for the clay one. The authors concluded that the adsorption of AMD was inversely proportional
361 to initial concentrations of acrylamide and remains insignificant when compared to the ability of
362 bacteria to breakdown acrylamide in natural environment. The low affinity of AMD to soil and clay
363 minerals in particular is likely due to its high hydrosolubility and small size (Sojka *et al.* 2007). Analysis
364 of a lake water, directly connected to groundwaters, suggest a likely transfer of AMD due to its
365 infiltration in soils (Junqua *et al.* 2013). The half-life of acrylamide monomer in rivers ranges from
366 weeks to months (Brown *et al.* 1980).

367

368 **4. Degradation of PAM-based flocculants**

369

370 ***4.1. Mechanisms of PAM degradation***

371 PAM-based flocculants degradation has been mainly studied for PAMs in aqueous solutions. The
372 degradation of PAMs results in a reduction of the molecular weight of the polymer and can adversely
373 affect the performance of such aqueous solutions in their commercial application as flocculants.
374 Degradation involving chain scission is accompanied by a significant change in the rheological
375 properties of the polymer solutions and this has often been used to monitor and express the extent of
376 the degradation. A correlation between the reduction in solution viscosity and performance as a
377 flocculating agent has been demonstrated in number of references. However the factors leading to
378 such degradation and the mechanisms of the degradation processes are not completely elucidated.

379 In environmental applications (such as in flocculation processes), very few experiments have been
380 conducted to quantify the PAM degradation in natural hydrosystems. Some reports have indicated that
381 polyacrylamide, in the presence of sunlight and glyphosate, photolytically degrades to acrylamide
382 monomer and this is a direct introduction of acrylamide into agricultural areas (Smith and Oehme
383 1993, Smith *et al.* 1996; Smith *et al.* 1997). The main difficulty encountered is due to the lack of simple
384 methods to analyze and quantify PAM in this context. Thus what is known about PAM degradation is
385 drawn from sparse reports, often using smaller molecular weight PAMs, or from indirect measurement
386 of decomposition. Most studies report slow degradation rates as a result of mechanical, photochemical
387 and biological processes.

388

389 ***Mechanical degradation***

390 Aqueous solutions of PAM or floc suspensions can both undergo mechanical degradation due to a
391 variety of processes such as stirring, pumping, injection and movement through porous media and
392 direct mechanical load. In the aggregate treatment plant, mechanical degradation of the PAM can
393 occur due to a shearing hydrodynamic field (mixing of the PAM-based flocculants with the process
394 waters and transportation of the sludge to tailings ponds along pipelines through pumping) or by direct
395 mechanical load (solid/liquid separation and sludge dewatering with a press-filter).

396 It was reported that a shearing hydrodynamic field induced chain bond scissions leading to a decrease
397 in the molecular weight of the polyacrylamide and then to a decrease in intrinsic viscosity of PAM
398 solutions (Paterson and Abernathy 1970; Abdel-Alim and Hamielec 1973; Nagashiro *et al.* 1975;
399 Povkh and Chernyuk 1986; Nguyen and Boger 1998; Avadiar *et al.* 2013). In particular, mechanical
400 degradation of polymers used for drag reduction in a turbulently flowing liquid has been studied by
401 several authors (Rho *et al.* 1996; Vlachogiannis *et al.* 2003; Pereira and Soares 2012). Losses in the
402 effectiveness of the polymers in reducing drag under flow were attributed either to fractures of the
403 PAM molecules leading to changes in the molecular weight distribution or to the fact that some or all of
404 the degradation could be associated with a decrease of aggregation in a shear flow (Wade and Kumar
405 1972; Vlachogiannis *et al.* 2003).

406 Regarding mechanical degradation of flocs, Tang *et al.* (2001) showed that the mechanical strength of
407 the floc depends on both the interparticle forces and on the floc density. In general, the rupture of floc
408 is classified either as “surface erosion” or as “large-scale fragmentation” (Yeung and Pelton 1996).
409 Erosion is the separation of small particles from the floc surface and it is attributed to shearing
410 stresses on the floc surface. Floc breakage during shear is suspected to involve the rupture of
411 hydrogen bonds (Senden *et al.* 2000; Taylor *et al.* 2002). Fragmentation refers to the break-up of flocs
412 into pieces of comparable size caused by pressure gradients across the entire body. Since flocs
413 break-up occurs at the weakest point in the floc structure, this usually results in more compact
414 aggregates, though smaller than the initial flocs. According to Caulfield (2002), it cannot be stated that
415 under the influence of a mechanical load polyacrylamides degrade to form their constituent monomeric
416 units such as acrylamide. Moreover, several authors reported that the flocs formed using
417 polyelectrolytes such as PAM are reformed after being broken up but do not regain their original size
418 and structure and form a new structure more compacted (Nguyen and Boger 1985; Leong *et al.* 1987;

419 Spicer *et al.* 1998). An example of experiments on mechanical degradation was performed by Rasteiro
420 (2008) who studied the degradation under hydrodynamic shearing induced by a recirculating pump of
421 two high molecular weight cationic PAMs used for the flocculation of precipitated calcium carbonate.
422 For the pump speeds tested, the flocs size decrease was attributed to floc erosion but it was also
423 stated that flocs reach another equilibrium state where aggregation and fragmentation rates are the
424 same. Moreover, it was reported that erosion may not depend on the flocculant concentration since it
425 depends on the interparticles bonds of the flocs; however erosion increased with the pump speed.
426 Reflocculation occurred only for the higher flocculant concentrations due to the excess of polymer and
427 most of the flocs resulting from the reflocculation process were observed to be more compact than the
428 original ones Rasteiro (2008).

429 Mechanical load can also induce enough shear-induced stress to cause bond scission and the
430 formation of a macromolecular radical (Caulfield 2002). For example, it was observed that filtration
431 under pressure of thickened suspensions from PAM flocculation of bauxite residue or brown coal lead
432 to significant breakdown in network structures and reduction in yield stresses (Nguyen and Boger
433 1985; Leong *et al.* 1987). The implementation of shear stresses that are higher than the initial yield
434 stresses of flocs suspension leads then to breakdown the intricacy involved within these complex
435 network structures (Avadiar *et al.* 2013).

436 Therefore, mechanical degradation of aqueous solutions of PAM or floc suspensions often involves
437 irreversible changes of the polymeric material including a marked reduction in viscosity and a scission
438 of the main chain bond and thus the generation of free radicals. However, none of the studies found in
439 the literature suggest that PAMs degrade to AMD under mechanical constraints.

440

441 ***Photodegradation of polyacrylamide***

442 PAM photodegradation is reported to be a free radical process that can lead to a cleavage of the
443 polymer backbone (bond scission) and the formation of lower molecular weight products (Decker
444 1989; Caulfield *et al.* 2002). The C-C, C-H and C-NH bonds in PAMs have bond strengths of 340, 420
445 and 414 kJ mol⁻¹ which can be cleaved by wavelengths of 325, 288 and 250 nm respectively (Crosby
446 1976). Even if the atmosphere absorbs most of the lower spectrum (less than approximately 300 nm),
447 it still leaves components from the UV and visible regions with sufficient energy to disrupt chemical
448 bonds (Rabek 1996). The free radicals formed from bond scission are extremely unstable and will

449 react with any suitable substrate. The reactions include combination to reform the saturated carbon-
450 carbon polymer backbone, disproportionation to form an unsaturated and saturated polymer chain end
451 and reaction with other entities present. The presence of oxygen in the system has a strong impact on
452 photodegradation of PAM as well as the impurities contained in the initial polymer.

453 Some authors have studied PAM degradation via advanced oxidation processes, such as Fenton
454 (Kurenkov *et al.* 2002), H₂O₂/UV (Caulfield *et al.* 2002), photo-Fenton (Giroto *et al.* 2008) and
455 O₃/H₂O₂/UV (Ren *et al.* 2006), but these reactions cannot occur in environmental conditions.

456 Most of the studies, dedicated to PAM photodegradation in environmental conditions, focus on the
457 potential release of AMD. This phenomenon was firstly observed by Smith *et al.* (1996, 1997), in
458 controlled laboratory conditions and under environmental conditions using an aqueous solution of a
459 PAM thickening agent (PAM mixed with glyphosate). However these results, their interpretation and
460 the overall experimental design raised some controversy, particularly the way that PAM was prepared
461 which could lead to incomplete dissolution of PAM (Kay-Schoemake *et al.* 1998a; Ver Vers 1999;
462 Caulfield *et al.* 2002). Ver Vers (1999) reported work using analogous degradation conditions but
463 yielding different results. This study points out problems in proper dilution of the emulsion and
464 attributes the detection of small amount of AMD over time to the gradual release of unreacted AMD
465 contaminants from the incompletely dissolved emulsion. The author concluded that PAM does not
466 degrade to AMD monomer in the presence of sunlight, even in presence of glyphosate. Kay-
467 Shoemake *et al.* (1998b) showed that the treatment of solutions of high molecular weight anionic
468 PAMs with UV radiations reduced the PAM chain length. They found also that the photodegraded
469 PAM samples did not support bacterial growth as sole source of carbon. As it is well recognized that
470 acrylamide is completely biodegradable (see section VI), these results suggest that AMD is not formed
471 during the photodegradation of the polyacrylamide. El-Mamouni *et al.* (2002) investigated the
472 photodegradation of a PAM solution (PAMs molecular weight ranging between 10⁵ and 10⁶ daltons)
473 exposed to UV irradiation (254 nm) prior to biomineralization. They observed a shift in the molecular
474 weight distribution, which demonstrates the degradation of the PAM chains into oligomers. A study
475 leaded by Caulfield *et al.* (2003) has reported that strong UV radiation at 254 nm released AMD from
476 solutions of a nonionic PAM. However, the release was very small. They also noted a reduction in
477 viscosity of the polymer solution, attributed to a lowering of the molecular weight consistent with
478 polymer chain scission. Suzuki *et al.* (1978, 1979) also reported a number of low molecular weight

479 PAM decomposition products when degraded using strong UV irradiation in the presence of ozone but
480 didn't notice any AMD monomer release. It must be noted that the studies of Kay-Schoemake *et al.*
481 (1998a), El Mamouni *et al.* (2002), Caulfield *et al.* (2003) and Suzuki *et al.* (1978, 1979) used UV
482 irradiations which are much more energetic than the sunlight's wavelengths. These studies cannot
483 confirm the attenuation of PAM by sunlight in natural environments.

484 Holliman *et al.* (2005) submitted samples of field-conditioned cross-linked PAM gels to a 365-nm UV
485 radiation in a daily light-dark cycle during 5 months without detectable acrylamide release. On the
486 other hand, they observed a gradual loss of gel properties they attribute to polymer chain scission
487 resulting in the production of PAM fragments. More recently, a study by Woodrow *et al.* (2008)
488 reported AMD release resulting from sunlight irradiation of aqueous polyacrylamide/iron mixtures. A
489 solution of commercial anionic PAM with different concentration of Fe^{3+} (from 0.02 to 4.4 ppm) was
490 submitted to UV irradiation in a simulator able to closely approximate solar irradiance in the
491 troposphere (equivalent to late summer noontime sunlight at 40° latitude – 300nm). Without Fe^{3+} , only
492 polymer chain scission was observed through HPLC measurement and decline of average molecular
493 weight. In the presence of Fe^{3+} , acrylamide release was also observed at acid/neutral pH whereas at
494 alkaline pH (~ 8.0), PAM/ Fe^{3+} remained stable under irradiation.

495 In conclusion, PAMs are susceptible to a wide range of photodegradative processes depending on the
496 reaction conditions. Photodegradation results in irreversible changes in both physical and chemical
497 properties of the PAMs, but the release of acrylamide still remains in question and would require
498 further clarification. Moreover photodegradation of PAM was not directly demonstrated in
499 environmental conditions in hydrosystems with field studies but only in laboratory aqueous solutions
500 without the presence of mineral particles.

501

502 **Biological degradation**

503 Whether they are used as flocculants for water clarification or in agricultural applications for the control
504 of soil erosion as well as in petrochemical productions to enhanced oil recovery, PAMs are in contact
505 with micro-organisms and can be subject to biodegradation. Most of the studies on the degradation of
506 PAM in agricultural soils concluded that the biodegradation of PAM did not result in the formation of
507 monomer (AMD) and that PAMs were generally resistant to microbial degradation (Caulfield *et al.*

508 2002). During these applications, the degradation seemed to be most likely due to environmental
509 factors such as UV light or chemical degradation (Seybold 1994).

510 The ability of bacteria to utilize PAM as sole nitrogen or sole carbon sources has been examined. Kay-
511 Shoemake *et al.* (1998a) demonstrated that PAM can be used as an organic nitrogen source for
512 microorganisms in soil. A bacterial growth was observed when cultures were produced on soil
513 samples with no other source of nitrogen. Intracellular and extracellular amidase activities were
514 detected in all the enrichment cultures able to use PAM as sole source of nitrogen and in soils treated
515 with PAM. Haveroen *et al.* (2005) also demonstrated that the addition of PAM stimulates microbial
516 activities and methanogenesis in anaerobic environments that are poor in nitrogen sources. In another
517 study, Kay-Shoemake *et al.* (1998b) show that indigenous bacteria, commonly found in soils, were
518 enabled to use PAM as sole carbon source. These results suggest that these microorganisms could
519 hydrolyse the amine group but were unable of cleaving the main carbon chain of the polymer.

520 PAMs are high molecular weight polymers and are unable to pass biological membranes, so the
521 biodegradation of these polymers requires the action of extracellular enzymes, like amidases, to
522 degrade the carbon chain. These characteristics limit the potential of biodegradation and can be at the
523 origin of the partial degradation of PAM (Caulfield *et al.* 2002). Few studies demonstrated the
524 biodegradation of these compounds. Nakamiya and Kinoshita (1995) were the first ones to report the
525 isolation of two PAM-degrading bacterial strains from soil samples. These strains, identified as
526 *Enterobacter agglomerans* A and *Azomonas macrocytogenes* B, were able to reduce the molecular
527 weight of a synthesized PAM and to degrade about 15 to 20% of the initial concentration of PAM in a
528 synthetic medium. Another study (Matsuoka *et al.* 2002) reported the isolation of two PAM-degrading
529 bacteria strains from soil, identified as *Bacillus sphaericus* n°2 and *Acinetobacter sp.* No. 11. These
530 strains grew on medium containing PAM as sole carbon and nitrogen sources and reduced,
531 respectively, by 16% and 19% of the initial PAM concentration. However, the degradation of PAM did
532 not result in the formation of AMD. As previously indicated, the biodegradation of PAM require
533 extracellular amidase activities. These enzymes seem to be enabled to liberate AMD directly from
534 PAM but can produce lower molecular weight polymers and can alter the nature of the substituents
535 attached to the polymer backbone (Caulfield *et al.* 2002, Kay-Shoemake *et al.* 1998b).

536 More recently, several bacterial strains isolated during oil recovery process have demonstrated the
537 ability to degrade Hydrolysed Polyacrylamide (HPAM) or PAM (Ma *et al.* 2010; Wen *et al.* 2010; Bao *et*

538 *al.* 2010). Ma *et al.* (2010) have isolated a *Clostridium bifermentans* H1 strain which can use HPMA as
539 sole carbon source and can hydrolyse the amine to carboxyl group by the molecule chain amine
540 hydrolysis. In their experimental conditions, the removing efficiency for HPAM reached 30.8 %. Wen *et*
541 *al.* (2010) reported the isolation of two strains of *Bacillus cereus* HWBI and *Bacillus flexu* HWBII from
542 activated sludge and oil-contaminated soil. Both strains were able to grow on a medium containing
543 PAM as sole carbon and nitrogen source and to degrade PAM at different rates. After 72 hours of
544 cultivation in a mineral salt medium (MSM) supplemented with 60 mg.L⁻¹ of PAM at 30°C, more than
545 70 % of the PAM was consumed and PAM was cleaved into smaller molecules. Another study
546 reported the isolation of two *Bacillus sp.* strains, named PM-2 and PM-3 from production water after
547 polymer flooding in an oilfield (Bao *et al.* 2010). The PAM removal efficiencies observed were lower
548 than others studies. After 7 days of cultivation at 40°C in a basal medium containing 0.5 g.L⁻¹ of HPMA
549 as a sole source of nitrogen, 36.3 % of the HPMA was degraded. Lower HPMA removal efficiencies
550 were observed when HPMA was used as sole carbon and nitrogen sources. These results suggested
551 that the PAM-degrading strains obtained more nutriment from the amide sides groups of HPAM using
552 amidase activities compared to cleaving the main carbon chain backbone (Kay-Shoemaker *et al.*
553 1998b).

554 Although the biodegradation of PAM has been studied with isolated strains in pure culture, in practical
555 work, the biodegradation of PAM requires interactions between several microbial species. Recently,
556 Liu *et al.* (2012) have described a PAM-degrading granular sludge which degraded PAM. The PAM-
557 degrading granules which were mostly composed by filamentous and bacilli bacteria used PAM as
558 sole carbon and nitrogen source and degrade PAM at a high rate. PAM concentration was decreased
559 from 66.7 mg.L⁻¹ to 32.9 mg.L⁻¹ in 24 hours but no AMD monomer was detected during the
560 biodegradation process. The main intermediate product was identified as polyacrylic acid with a low
561 molecular weight.

562 These studies presented in this review demonstrated that PAM can be subjected to microbial
563 degradation. PAM was principally used as a nitrogen source by hydrolysis of the amine groups from
564 the polymer. The presence of glucose at low concentrations which can be used as co-metabolism
565 substrate can improve the PAM degradation but the biodegradation efficiency seems to be affected by
566 the initial concentration of PAM (Wen *et al.* 2010). The biodegradation of PAM induced changes in the
567 structure of the polymer (reduction of the molecular weight) but no evidence of release of AMD

568 monomer has been found. Although amidases are involved in the mechanism of degradation of PAM,
569 the reaction pathway for the microbial degradation of PAM is not clearly identified.

570

571 **4.2. Mechanisms of acrylamide degradation**

572 ***Bio-degradation of acrylamide***

573 AMD has an inhibitory effect on sulfhydryl proteins and hence on growth of microorganisms (Cavins
574 and Friedman 1968). However, some microorganisms are able to grow in the presence of AMD and
575 degrade this molecule. The microbial degradation of AMD has been demonstrated for several decades
576 in studies investigating the behavior, fate and impacts of AMD in natural environments amended with
577 PAMs. Microbial degradation was shown to occur in soils (Shanker *et al.* 1990, Shukor *et al.* 2009a),
578 sediments (Labahn *et al.* 2010), wastewater (Buranasilp and Charoenpanich 2011, Nawaz *et al.* 1996)
579 and hydrosystems (Labahn *et al.* 2010) and is mediated by microorganisms commonly presents in
580 these environments. Most studies deal with the biodegradation of AMD by bacteria but some fungi
581 have also been shown to degrade AMD. In particular, filamentous fungi such as some strains of
582 *Aspergillus oryzae* have interesting AMD degradation ability (Wakaizumi *et al.* 2009).

583 Among bacteria having an AMD degrading ability, most documented belong to species within the
584 genera *Bacillus* (Shukor *et al.*, 2009a), *Pseudomonas* (Labahn *et al.* 2010, Sathesh Prabu and
585 Thatheyus 2007, Shanker *et al.* 1990, Shukor *et al.* 2009b) and *Rhodococcus* (Nawaz *et al.* 1998).
586 However it is now establish that this activity is more widespread among bacteria such as *Enterobacter*
587 *aerogenes* (Buranasilp and Charoenpanich 2011), *Enterococcus faecalis* (Buranasilp and
588 Charoenpanich 2011) *Klebsiella pneumoniae* (Nawaz *et al.* 1996), *Burkholderia* sp. , *Xanthomonas*
589 *maltophilia* (Nawaz *et al.* 1993), *Kluyvera georgiana* (Thanyacharoen *et al.* 2012), *Variovorax*
590 *boronicumulans* (Liu ZH. *et al.* 2013), *Geobacillus thermoglucosidasius* (Cha and Chambliss 2013),
591 *Ralstonia eutropha* (Cha and Chambliss 2011) or *Arthrobacter* strains (Yamada *et al.* 1979). The
592 concentration of AMD that can be tolerated and degraded depends on the bacterial strain. As an
593 example and among strains that tolerate the higher concentration of AMD, *Bacillus cereus* is able to
594 degrade and assimilate AMD at concentrations as high as 3 g. L⁻¹ (Shukor *et al.* 2009a). Shanker *et*
595 *al.* (1990) isolated a *Pseudomonas* sp. strain able to degrade 4 g. L⁻¹ AMD, and *Enterobacter*
596 *aerogenes* was shown to grew well in the presence of 5 g. L⁻¹ AMD (Buranasilp and Charoenpanich
597 2011). It has also been demonstrated that bacteria able to degrade AMD are also able to degrade

598 other aliphatic amides such as acetamide, urea, propionamide (Buranasilp and Charoenpanich 2011;
599 Shukor *et al.* 2009a).

600 The biodegradation of AMD was first shown to be aerobic but recent studies have demonstrated the
601 involvement of some bacteria in the anaerobic biodegradation of AMD. In particular, nitrate reducers
602 have been shown to significantly remove AMD (Labahn *et al.* 2010). These authors also demonstrated
603 the potential of sulfate- and iron-reducers to degrade AMD. The biodegradation of AMD leads to the
604 production of acrylic acid (AA) and ammonia (NH₃). This could explain that AMD degrading
605 microorganisms use AMD as nitrogen source and sometimes also as carbon source, and that some
606 bacteria can grow as AMD as the sole sources of N and C (Shukor *et al.* 2009a, Nawaz *et al.* 1994,
607 1996). From an experimental point of view, AMD can be analyzed directly either by HPLC or by UPLC
608 depending on the concentration (Togola *et al.*, 2014; Guezennec *et al.*, 2014; Touze *et al.*, 2014;
609 VerVers, 1999). The degradation of AMD can also be easily monitored by the quantification of
610 ammonia that can be achieved using indicator such as Phenol red that changes in color (from red to
611 pink) in the presence of ammonia (Santoshkumar *et al.* 2010).

612 Many studies also underlined that the AMD biodegradation activity of free cells and immobilized cells
613 can be different. It is in fact usually admitted that immobilized bacterial technology offers several
614 advantages over the application of free cells in the degradations of pollutants (Nawaz *et al.* 1998):
615 immobilized cells are known to retain high catabolic activity, can be reused, efficiently mineralize
616 substrates and offers physical protection from predators and parasites. Nawaz *et al.* (1998) thus
617 demonstrated that immobilized cells of *Rhodococcus sp.* rapidly degraded 64 and 128 mM AMD in 3
618 and 5 hours respectively whereas free cells took more than 24 h to degrade 64 mM AMD. Moreover
619 an AMD concentration of 128 mM inhibited the growth of the free cells. Sathesh Prabu and Thatheyus
620 (2007) observed that free *Pseudomonas aeruginosa* cells began the degradative activity of AMD only
621 after incubation of 24 h, while immobilized cells started before 24 h. On the opposite, Buranasilp and
622 Charoenpanich (2011) showed that AMD degradation by immobilized cells of *Enterobacter aerogenes*
623 was slower than that of free cells. This could be explained by an inappropriate immobilization process
624 or difficulties in substrate-cell contact.

625 In addition to the state (free or immobilized) of cells, the biodegradation of AMD is influenced by
626 several physical factors such as AMD concentration, temperature, pH, the presence of metals. Nawaz
627 *et al.* (1998) demonstrated that the biodegradation of *Rhodococcus sp.* was lower at low temperature

628 (less than 60% of AMD was degraded at 10°C in 4 h whereas 100 % of AMD was degraded in less
629 than 3 h at 28°C and 45°C). An incubation temperature of 55°C completely inhibited the degradation of
630 AMD by this strain. pH 7 favored maximum degradation of AMD, and activity was slower at pH 8 and
631 very little degradation was measured at pH 6. These authors also demonstrated that nickel and cobalt
632 slowed the AMD degradation ability (when used at 5 mM) or inhibit (when used at 10 mM) the AMD
633 degradation ability. The highest rate of degradation of AMD was observed in the presence of iron but
634 iron concentrations higher than 10 mM inhibited AMD degradation. Sathesh Prabu and Thatheyus
635 (2007) showed that mercury and chromium inhibited AMD degradation by *Pseudomonas aeruginosa*
636 while nickel enhanced the process.

637

638 ***AMD biodegradation in environmental conditions***

639 If numerous authors have investigated the mechanisms of AMD biodegradation and the
640 microorganisms involved, very few have quantified AMD biodegradation rate and kinetics in
641 environmental conditions (temperature, AMD concentration, natural samples...). Those study exhibit
642 contrasting results (see Table 1) and are difficult to compare since most of biodegradation tests have
643 been conducted in various range of AMD concentration. The work carried out by Labahn *et al.* (2010)
644 on natural water and slurry samples spiked with 50 µg.L⁻¹ AMD show the impact of the temperature on
645 AMD biodegradation: at 15°C, no degradation is detected after 72h whereas at 25°C, AMD
646 degradation is complete after 24h. These results confirm those obtained by Nawaz *et al.* (1998) in
647 controlled laboratory biodegradation study using pure strain of *Rhodococcus* sp (see section above).
648 Croll (1974) investigated AMD biodegradation in river water samples spiked with 10 µg.L⁻¹ AMD. One
649 of the water samples was inoculated with a culture capable of degrading acrylamide. In this case,
650 AMD degradation was complete after 50h, the lag period being 5h. In water samples which were not
651 inoculated, the time required to fully degrade AMD reached more than 300h with a lag period of 50h.
652 Brown (1979) used also river water samples but with higher concentration of AMD (5 mg.L⁻¹) and
653 investigated the influence of light on the biodegradation. In both cases (with and without light), the
654 results were the same: AMD was fully degraded after 125h with a lag time of 75h. In a more recent
655 study Ver Vers (1999) demonstrated that the time required for complete AMD biodegradation could
656 vary from 480h to more than 1100h according to the type of water samples (tap, river and lake). For

657 the author, those differences could indicate that some species in the water may be interacting with
658 AMD, either accelerating or hindering monomer degradation.

659 The results obtained by these author show that AMD biodegradation is probably influenced by
660 different types of factors such as the initial AMD concentration, the nature of the media in which the
661 degradation occurs, its microbial composition... Given the conditions used, the degradation can be
662 quick but the time required for a complete degradation of AMD is never lower than 1 day. In some
663 cases several days are needed to degrade the monomer.

664

665 ***Metabolism of acrylamide and enzymes involved in bio-degradation***

666 As mentioned before, AMD can be degraded by microorganisms under aerobic and anaerobic
667 condition, and serves as nitrogen and/or carbon sources. Despite the fact that microbial degradation of
668 acrylamide has been explored extensively, the metabolic pathways of degradation are unclear. The
669 microbial degradation of AMD is initially due to the presence of an enzyme named acylamide
670 amidohydrolase (E.C. 3.5.1.4) or amidase. Amidases catalyze the hydrolysis of an amide to free
671 carboxylic acids and free ammonium. In the case of acrylamide, the AMD is deaminated to ammonia
672 and acrylic acid (AA). These enzymes belong to the family of hydrolases which acts on carbon-
673 nitrogen bonds other than peptide bonds. They are ubiquitous enzymes that can have various
674 biological functions and the substrate preferences differ. In their review, Sharma *et al.* (2009) widely
675 described these enzymes, their uses in industries (and their potential applications in the production of
676 various chemicals), their structural and biochemical properties, substrates and some purification
677 protocols.

678 Amidase activity has been identified in various bacteria genus, commonly found in soils, including
679 *Rhodococcus*, *Bacillus*, *Brevibacterium*, *Alcaligenes* and *Pseudomonas* and in fungi (Kay-Shoemake
680 *et al.* 1998b, Wakaizumi *et al.* 2009). These micro-organisms are able to produce a single amidase or
681 numerous isoenzymes during growth with amide substrate. Furthermore, amidases exhibited
682 enzymatic activity with relatively limited substrate specificity while others are able to hydrolysis a broad
683 range of spectrum. Among amidases, some have been demonstrated to be involved in the first step of
684 the AMD-degrading microbial pathways for the catalytic hydrolysis of AMD to acrylic acid and
685 ammonia (Sharma *et al.* 2009).

686 Although some amidases are constitutive (Nawaz *et al.* 1994), microbial amidases are generally
687 inducible. It has been demonstrated that amides and AMD can be an inducer of amidase expression
688 and some carbon sources as glucose or succinate can induce a catabolic repression (Potts and Clarke
689 1976, Sharma *et al.* 2009). The regulation of amidase (encoded by *amiE* gene) expression has been
690 study in *Pseudomonas aeruginosa*. In this microorganism, the induction of amidase synthesis is
691 positively regulated by a regulatory protein encoded by *amiR* and a second regulatory gene, *amiC*,
692 which negatively regulates amidase expression (Clarke *et al.* 1981, Cousens *et al.* 1987, Wilson. and
693 Drew 1991, Wilson *et al.* 1993, Wilson and Drew 1995). However, under non-inducing condition, low
694 levels of amidase activity are always present within the cells. But, the cellular mechanism for induction
695 or sensing remains unclear because the PAM-polymer is too large to enter the cells and to act as a
696 direct inducer. Shanker *et al.* (1990) have demonstrated that addition of glutamate or ammonium
697 sulfate as additional nitrogen source induced increase in degradation potential.

698 Several amidases using AMD as substrate have been purified from AMD-degrading bacteria (Cha and
699 Chambliss 2013; Egorova *et al.* 2004; Lavrov *et al.* 2010; Liu *et al.* 2013; Makhongela *et al.* 2007;
700 Nawaz *et al.* 1994; Nawaz *et al.* 1996; Ohtaki *et al.* 2010; Syed *et al.* 2012). The amidases purified
701 from AMD-degrading bacteria are mainly monomer but that from *Burkholderia sp.* is a homotetramer
702 (Syed *et al.* 2012). The molecular weight, pH and temperature optima depend on the enzyme (see
703 Table 2). It has been also demonstrated that the catalytic hydrolysis of AMD by amidase can be
704 inhibited or enhanced by metals (Nawaz *et al.* 1994). These authors showed that amidase activity of
705 *Rhodococcus sp.* was enhanced by Fe^{2+} , Ba^{2+} and Cr^{2+} but not dependent on these cations. On the
706 other side, the enzyme activity was partially inhibited by Mg^{2+} and totally inhibited in the presence of
707 Ni^{2+} , Hg^{2+} , Cu^{2+} and Co^{2+} .

708 The biodegradation of AMD requires not only amidase but also enzymes able to metabolize the
709 intermediate products. Indeed, the biodegradation of AMD induces the production of ammonia and
710 acrylic acid. However, the subsequent fate of these products is not clear but probably involves specific
711 catabolic pathways. For AMD degrading-bacteria, AA is probably reduced to generate energy for
712 bacterial growth but the catabolic pathway is not clear. Wampler and Ensign (2005) suggested that AA
713 is degraded to β -hydropropionate via hydroxylation reaction, then oxidized to CO_2 or reduced to
714 propionic acid. Recently, Charoenpanich and Tani (2014) have studied the response of an acrylamide-
715 degrader *Enterococcus aerogenes* to AMD using proteome analysis. These authors have identified

716 fifteen proteins differentially expressed after growth in presence of AMD and have proposed a model
717 of metabolic pathway of AMD degradation based on these results (see Fig. 3).

718 Finally, the identification of a “universal” pathway and the enzymes involved in AMD metabolism in
719 prokaryotes remains a challenge for the optimization of the bioremediation of AMD.

720

721 **5. Conclusion**

722 This review summarizes the literature which gives information and scientific data on the fate of PAM-
723 based flocculants in hydrosystems. PAM and associated AMD have been considered and three main
724 families of phenomenon have been investigated: transport, adsorption and degradation. Most of the
725 information gathered in this review was derived from studies conducted in the frame of PAM use for
726 agricultural application. Due to PAM ability to adsorb onto mineral particles, its transport in surface
727 water, groundwater and soils is rather limited and restricted to specific conditions. PAM adsorption is
728 rapid and irreversible, and desorption occurs only at a very small extent. In the contrary, the
729 adsorption of acrylamide onto particles is very low, which is likely due to its high hydrosolubility and
730 small size. This low affinity to soil and clay minerals could favor its transfer in surface waters and
731 ground water, but very few studies reports AMD analysis in natural water. Moreover, several studies
732 have demonstrated microbial degradation of acrylamide in environments amended with
733 polyacrylamide. The biodegradation of acrylamide could thus limit its transfer in the environment.
734 PAMs are high molecular weight polymers and are unable to pass biological membranes, so the
735 biodegradation of these polymers requires the action of extracellular enzymes, like amidases, to
736 degrade the carbon chain. These characteristics limit the potential of PAM biodegradation. PAM can
737 be also subject of photodegradation and mechanical degradation. In any case, most of the studies
738 report slow degradation rates without AMD release. The main conclusion of this review is that, despite
739 the abundant literature dedicated to PAM use as flocculants, there is an obvious lack of pertinent and
740 detailed data about the behavior, the fate and the impact of PAM-based flocculants in environments.
741 Lot of work has still to be carried out to elucidate the mechanisms of degradation processes as well as
742 to quantify the transfer of PAM and AMD in hydrosystems.

743

744 **Acknowledgments:** The research leading to this publication has received funding from the ANR in
745 the framework of AquaPOL project (CES programme, project n°1443).

746

747 **References**

748

749 Abdel-Alim AH, Hamielec AE (1973) Shear degradation of water-soluble polymers. I. Degradation of
750 polyacrylamide in a high-shear couette viscometer. *J. Appl. Polym. Sci.* 17:3769–3778

751 Aly SM, Letey J (1988) Polymer and water-quality effects on flocculation of montmorillonite. *Soil*
752 *Science Society of America Journal* 52:1453-1458

753 Andersen, F.A. 2005. Amended final report on the safety assessment of polyacrylamide and
754 acrylamide residues in cosmetics, *International Journal of Toxicology*, 24:21-50.

755 Arrowood TJ (2008) Determining the Fate and Transport of the Acrylamide Monomer (AMD) in Soil
756 and Groundwater Systems, University of Nevada, University of Nevada at Las Vegas, Las
757 Vegas, NV, United States (USA), 132 pp

758 Avadiar L, Leong Y, Fourie A, Nugraha T (2013) Behaviours of Kaolin Suspensions with Different
759 Polyacrylamide (PAM) Flocculants under Shear. Chemeca 2013, Barton, ACT, 2013

760 Bao M, Chen Q, Li Y, Jiang G (2010) Biodegradation of partially hydrolyzed polyacrylamide by
761 bacteria isolated from production water after polymer flooding in an oil field. *J. Hazard. Mater.*
762 184:105-110

763 Barvenik FW (1994) Polyacrylamide characteristics related to soil applications. *Soil Science* 158:235-
764 243

765 Benhur M, Faris J, Malik M, Letey J (1989) Polymers as soil conditioners under consecutive irrigations
766 and rainfall. *Soil Science Society of America Journal* 53: 1173-1177

767 Benhur M, Letey J (1989) Effect of polysaccharides, clay dispersion, and impact energy on water
768 infiltration. *Soil Science Society of America Journal* 53:233-238

769 Benhur M, Malik M, Letey J, Mingelgrin U (1992) Adsorption of polymers on clays as affected by clay
770 charge and structure, polymer properties, and water-quality. *Soil Science* 153:349-356

771 Bicerano J (1994) Predicting key polymer properties to reduce erosion in irrigated soil. *Soil Science*
772 158 :255-266

773 Bottero, Lartiges (1992) Séparation liquide/solide par coagulation-floculation : les
774 coagulants/floculants, mécanismes d'agrégation, structures et densité des floccs. *Mines et*
775 *Carrières* 4

776 Bouranis DL, Theodoropoulos AG, Drossopoulos JB (1995) Designing synthetic-polymers as soil
777 conditioners. *Communications in Soil Science and Plant Analysis* 26 :1455-1480

778 Bouranis DL (1998) Designing synthetic soil conditioners via postpolymerization reactions. *Handbook*
779 *of Soil Conditioners*, 333-362

780 Brown L, Rhead M (1979) Liquid-chromatographic determination of acrylamide monomer in natural
781 and polluted aqueous environments. *Analyst* 104:391-399

782 Brown L, Bancroft KCC, Rhead MM (1980) Laboratory studies on the adsorption of acrylamide
783 monomer by sludge, sediments, clays, peat and synthetic resins. *Water Research* 14:779-781

784 Buranasilp K, Charoenpanich J (2011) Biodegradation of acrylamide by *Enterobacter aerogenes*
785 isolated from wastewater in Thailand. *J Environ Sci (China)* 23:396-403

786 Callebaut F, Gabriels D, De Boodt M (1979) The effect of polymer structure on soil physico-chemical
787 properties and soil water evaporation. *Journal of Chemical Technology and Biotechnology*.
788 29(12):723–729

789 Caulfield MJ, Qiao GG, Solomon DH (2002) Some aspects of the properties and degradation of
790 polyacrylamides. *Chem Rev* 102:3067-84

791 Caulfield MJ, Hao X, Qiao GG, Solomon DH (2003) Degradation on polyacrylamides. Part I. Linear
792 polyacrylamide. *Polymer* 44:1331-1337

793 Cavins JF, Friedman M (1968) Specific modification of protein sulfhydryl groups with alpha,beta-
794 unsaturated compounds. *J Biol Chem* 243:3357-60

795 Cha M, Chambliss GH (2011) Characterization of Acrylamidase isolated from a newly isolated
796 acrylamide-utilizing bacterium, *Ralstonia eutropha* AUM-01. *Curr. Microbiol.* 62:671-678

797 Cha M, Chambliss GH (2013) Cloning and sequence analysis of the heat-stable acrylamidase from a
798 newly isolated thermophilic bacterium, *Geobacillus thermoglucosidasius* AUT-01.
799 *Biodegradation* 24:57-67

800 Chamberlain P, Cole R (1996) Influence of structure and formulation on the efficacy of
801 polyacrylamides as soil stabilizers, *Managing Irrigation-Induced Erosion and Infiltration with*
802 *Polyacrylamide*, College of Southern Idaho, Twin Falls, ID, USA, pp. 83-87

803 Chang SH, Ryan ME, Gupta RK, Swiatkiewicz B (1991) The adsorption of water soluble polymers on
804 mica, talc limestone, and various clay minerals. *Colloids and Surfaces* 59 :59-70

805 Charoenpanich, J and Tani, A. (2014). Proteome Analysis of Acrylamide-induced Proteins in a Novel
806 Acrylamide-degrader *Enterobacter aerogenes* by 2D Electrophoresis and MALDI-TOF-MS.
807 CMU. J. Nat. Sci. 13:11.

808 Chiappa L, Mennella A, Lockhart TP, Burrafato G (1999) Polymer adsorption at the brine/rock
809 interface: the role of electrostatic interactions and wettability. *Journal of Petroleum Science*
810 *and Engineering* 24:113-122

811 Clarke PH, Drew RE, Turberville C, Brammar WJ, Ambler RP, Auffret AD (1981) Alignment of cloned
812 *amiE* gene of *Pseudomonas aeruginosa* with the N-terminal sequence of amidase. *Biosci Rep*
813 1:299-307

814 Cousens DJ, Clarke PH, Drew R (1987) The amidase regulatory gene (*amiR*) of *Pseudomonas*
815 *aeruginosa*. *J Gen Microbiol* 133:2041-52

816 Croll BT, Arkell GM, Hodge RPJ (1974) Residues of acrylamide in water. *Water Research* 8, 989-993

817 Crosby DG (1976): Herbicide photochemical decomposition. In: Kearney PCaK, D.D. (Editor),
818 *Berbicides : Chemistry Degradation and Mode of Action*. Marcel Dekker Inc., New York, pp.
819 836-841

820 de Rosemond SJC, Liber K (2004) Wastewater treatment polymers identified as the toxic component
821 of a diamond mine effluent. *Environ. Toxicol. Chem.* 23:2234– 2242.

822 Decker C (1989) Effect of UV radiation polymers. In: Cheremisinoff NP (Editor), *Handbook of Polymer*
823 *Science and Technology*. Marcel Dekker Inc., New York, pp. 541-608

824 Deng YJ, Dixon JB, White GN, Loeppert RH, Juo ASR (2006) Bonding between polyacrylamide and
825 smectite. *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 281:82-91

826 Egorova K, Trauthwein H, Verseck S, Antranikian G (2004) Purification and properties of an
827 enantioselective and thermoactive amidase from the thermophilic actinomycete
828 *Pseudonocardia thermophila*. *Appl. Microbiol. Biotechnol.* 65:38-45

829 El-Mamouni R, Frigon JC, Hawari J, Marroni D, Guiot S (2002) Combining photolysis and
830 bioprocesses for mineralization of high molecular weight polyacrylamides. *Biodegradation*
831 13:221-227

832 ElHardy OA, Abd ElHardy BM (1989) The interaction between polyacrylamide as a conditioner for
833 sandy soils and some plant nutrients. I. Effect on the mechanical strength and stability and
834 soil structure. *Egyptian Journal of Soil Science* 219:51-56

835 Entry JA, Sojka RE, Watwood M, Ross C (2002) Polyacrylamide preparations for protection of water
836 quality threatened by agricultural runoff contaminants. *Environ Pollut* 120:191-200

837 Entry JA, Sojka RE, Hicks BJ (2008) Carbon and nitrogen stable isotope ratios can estimate anionic
838 polyacrylamide degradation in soil. *Geoderma* 145:8-16

839 EPA US (2010) Toxicological Review of Acrylamide in Support of Summary Information on the
840 Integrated Risk Information Systems (IRIS), U.S. Environmental Protection Agency

841 European Parliament (1999) Directive 1999/45/EC concerning the approximation of the laws,
842 regulations and administrative provisions of the Member States relating to the classification,
843 packaging and labelling of dangerous preparations. *Official Journal of the European Union*
844 L200/1. European Union, Brussels

845 European Parliament (2006) Regulation 1907/2006/EC concerning the Registration, Evaluation,
846 Authorisation and Restriction of Chemicals (REACH). *Official Journal of the European Union*
847 L396/1. European Union, Brussels

848 European Union (2002). Risk assessment report on acrylamide. CAS N°: 79-06-1 (<http://europa.eu.int>)

849 Fitzpatrick CSB, Fradin E, Gregory J (2004) Temperature effects on flocculation, using different
850 coagulants. *Water Science and Technology* 50:171-175

851 Giroto JA; Teixeira AC, Nascimento CA *et al.* (2008) Photo-Fenton removal of water-soluble polymers.
852 *Chemical Engineering and Processing*. 47 2361-2369

853 Goodrich MS, Dulak LH, Friedman MA, Lech JJ (1991) Acute and long-term toxicity of water-soluble
854 cationic polymers to rainbow-trout (*oncorhynchus-mykiss*) and the modification of toxicity by
855 humic-acid. *Environmental Toxicology and Chemistry* 10:509-515

856 Graveling GJ, Ragnarsdottir KV, Allen GC, Eastman J, Brady PV, Balsley SD, Skuse DR (1997)
857 Controls on polyacrylamide adsorption to quartz, kaolinite, and feldspar. *Geochimica Et*
858 *Cosmochimica Acta* 61:3515-3523

859 Greenlan DJ (1963) Adsorption of polyvinyl alcohols by montmorillonite. *Journal of colloid science*
860 18:647-664

861 Guezennec AG, Michel C, Desroche N, Togola A, Touzé S (Submitted 2014) Microbial aerobic and
862 anaerobic degradation of acrylamide in sludges and industrial and natural waters - sand and
863 gravel quarry case study. *Environmental Science and Pollution Research (SI AQUAPOL)*

864 Hamilton M, Reinert K, Freeman MB (1994) Aquatic risk assessment of polymers. Environmental
865 Science and Technology 28:187A-192A

866 Hanson AT, Cleasby JL (1990) The effect of temperature on turbulent flocculation – fluid-dynamics
867 and chemistry. Journal American Water Works Association 82:56-73

868 Harford AJ, Hogan AC, Jones DR, van Dam RA (2011) Ecotoxicological assessment of a
869 polyelectrolyte flocculant. Water Research 45: 6393-6402

870 Haschke H, Miles MJ, Sheppard S (2002) Adsorption of individual polyacrylamide molecules studied
871 by atomic force microscopy. Single Molecules 3:171-172

872 Haveroen ME, MacKinnon MD, Fedorak PM (2005) Polyacrylamide added as a nitrogen source
873 stimulates methanogenesis in consortia from various wastewaters. Water Res 39:3333-41

874 Heath AR, Bahri PA, Fawell PD, Farrow JB (2006) Polymer flocculation of calcite: Experimental results
875 from turbulent pipe flow. AIChE Journal 52:1284-1293

876 Hollander AF, Somasundaran P, Gryte CC (1981) Adsorption characteristics of polyacrylamide co-
877 polymers on sodium kaolinite. Journal of Applied Polymer Science 26:2123-2138

878 Holliman PJ, Clark JA, Williamson JC, Jones DL (2005) Model and field studies of the degradation of
879 cross-linked polyacrylamide gels used during the revegetation of slate waste. Science of the
880 Total Environment 336:13-24

881 Inyang HI, Bae S (2005) Polyacrylamide sorption opportunity on interlayer and external pore surfaces
882 of contaminant barrier clays. Chemosphere 58:19-31

883 Jin RR, Hu WB, Hou XJ (1987) Mechanism of selective flocculation of hematite from quartz with
884 hydrolyzed polyacrylamide. Colloids and Surfaces 26:317-331

885 Junqua G, Spinelli S, Gonzalez C, (2013) Occurrence and fate of acrylamide in water-recycling
886 systems and sludge in aggregate industries Environmental Science and Pollution Research *in*
887 *press* doi:10.1007/s11356-014-3022-5

888 Kay-Shoemake JL, Watwood ME, Lentz RD, Sojka RE (1998a) Polyacrylamide as an organic nitrogen
889 source for soil microorganisms with potential effects on inorganic soil nitrogen in agricultural
890 soil. Soil Biology and Biochemistry 30:1045-1052

891 Kay-Shoemake JL, Watwood ME, Sojka RE, Lentz RD (1998b) Polyacrylamide as a substrate for
892 microbial amidase in culture and soil. Soil Biology and Biochemistry 30:1647-1654

893 Kurenkov VF, Hartan HG, Lobanov FI (2002) Degradation of polyacrylamide and its derivatives in
894 aqueous solutions. Russian Journal of Applied Chemistry. 75:1039-1050

895 Labahn SK, Fisher JC, Robleto EA, Young MH, Moser DP (2010) Microbially Mediated Aerobic and
896 Anaerobic Degradation of Acrylamide in a Western United States Irrigation Canal. Journal of
897 Environmental Quality 39:1563-1569

898 Laird DA (1997) Bonding between polyacrylamide and clay mineral surfaces. Soil Science 162:826-
899 832

900 LaMer VK, Healy TW (1963) Adsorption-flocculation reactions of macromolecules at the solid-liquid
901 interface. Review of Pure and Applied Chemistry 13:112-132

902 Lande SS, Bosch SJ, Howard PH (1979) Degradation and leaching of acrylamide in soil. J. Environ.
903 Qual. 8:133-137

904 Lavrov KV, Zalunin IA, Kotlova EK, Yanenko AS (2010) A new acylamidase from *Rhodococcus*
905 erythropolis TA37 can hydrolyze N-substituted amides. Biochemistry (Mosc) 75:1006-1013

906 Lecourtier J, Lee LT, Chauveteau G (1990) Adsorption of polyacrylamide on siliceous minerals.
907 Colloids and Surfaces 47:219-231

908 Lee JJ, Fuller GG (1985) Adsorption and desorption of flexible polymer-chains in flowing systems.
909 Journal of Colloid and Interface Science 103:569-577

910 Lee LT, Somasundaran P (1991) Effect of inorganic and organic additives on the adsorption of
911 nonionic polyacrylamide on hematite. Journal of Colloid and Interface Science 142:470-479

912 Lentz RD, Sojka RE (2000) Applying polymers to irrigation water: Evaluating strategies for furrow
913 erosion control. Transactions of the Asae 43:1561-1568

914 Lentz RD, Sojka RE, Mackey BE (2002) Fate and efficacy of polyacrylamide applied in furrow
915 irrigation: Full-advance and continuous treatments. Journal of Environmental Quality 31:661-
916 670

917 Leong YK, Creasy DE, Boger DV, Nguyen QD (1987) Rheology of brown coal-water suspensions.
918 Rheologica Acta 26:291-300

919 Liber K, Weber L, Levesque C (2005) Sublethal toxicity of two wastewater treatment polymers to lake
920 trout fry (*Salvelinus namaycush*). Chemosphere 61:1123-1133

921 Liu L, Wang Z, Lin K, Cai W (2012) Microbial degradation of polyacrylamide by aerobic granules.
922 Environ Technol 33:1049-54

923 Liu ZH, Cao YM, Zhou QW, Guo K, Ge F, Hou JY, Hu SY, Yuan S, Dai YJ (2013) Acrylamide
924 biodegradation ability and plant growth-promoting properties of *Variovorax boronicumulans*
925 CGMCC 4969. Biodegradation In Press

926 Lu JH, Wu L, Letey J (2002) Effects of soil and water properties on anionic polyacrylamide sorption.
927 Soil Science Society of America Journal 66:578-584

928 Lu JH, Wu LS (2013) Polyacrylamide distribution in columns of organic Matter-Removed Soils
929 following Surface Application. J. Environ. Qual. 32:674-680

930 Lurie M, Rebhun M (1997) Effect of properties of polyelectrolytes on their interaction with particulates
931 and soluble organics. Water Science and Technology 36:93-101

932 Lyklema J, Flerer GJ (1987) Electrical contributions to the effect of macromolecules on colloid stability.
933 Colloids and Surfaces 25:357-368

934 Ma Y, Yu H, Pan W, Liu C, Zhang S, Shen Z (2010) Identification of nitrile hydratase-producing
935 *Rhodococcus ruber* TH and characterization of an *amiE*-negative mutant. Bioresour. Technol.
936 101:285-291

937 McCollister DD, Owen F & Rowe VK (1964) Toxicology of acrylamide. Toxicol. Appl. Pharmacol.
938 6:172-181

939 Makhongela HS, Glowacka AE, Agarkar VB, Sewell BT, Weber B, Cameron RA, Cowan DA, Burton
940 SG (2007) A novel thermostable nitrilase superfamily amidase from *Geobacillus pallidus*
941 showing acyl transfer activity. Appl. Microbiol. Biotechnol. 75:801-811

942 Malik M, Letey J (1991) Adsorption of polyacrylamide and polysaccharide polymers on soil materials.
943 Soil Science Society of America Journal 55:380-383

944 Malik M, Nadler A, Letey J (1991) Mobility of polyacrylamide and polysaccharide polymer through soil
945 materials. Soil technology 4:255-263

946 Matsuoka H, Ishimura F, Takeda T, Hikuma M (2002) Isolation of polyacrylamide-degrading
947 microorganisms from soil. Biotechnology and Bioprocess Engineering 7:327-330

948 Michaels AS, Morelos O (1955) Polyelectrolyte adsorption by kaolinite. Industrial and Engineering
949 Chemistry 47, 1081-1809

950 Molak, V. Acrylamide: A Review of the Literature in NIOH and NIOSH Basis for an Occupational
951 Health Standard. US Department of Health and Human Services, 1991.

952 Mortland (1970) Clay-organic complexes and interactions. Advances in Agronomy 22:75-117

953 Mpofu P, Addai-Mensah J, Ralston J (2004) Temperature influence of nonionic polyethylene oxide and
954 anionic polyacrylamide on flocculation and dewatering behavior of kaolinite dispersions.
955 Journal of Colloid and Interface Science 27: 145-156

956 Nabzar L, Pefferkorn E, Varoqui R (1984) Polyacrylamide sodium kaolinite interactions – flocculation
957 behavior of polymer clay suspensions. Journal of Colloid and Interface Science 102:380-388

958 Nabzar L, Pefferkorn E, Varoqui R (1988) Stability of polymer clay suspensions – the polyacrylamide
959 sodium kaolinite system. Colloids and Surfaces 30:345-353

960 Nadler A, Letey J (1989) Adsorption-isotherms of polynions on soils using tritium labeled compounds.
961 Soil Science Society of America Journal 53:1375-1378

962 Nadler A, Malik M, Letey J (1992) Desorption of polyacrylamide and polysaccharide polymers from
963 soil materials. Soil Technology 5:91-95

964 Nagashiro W, Tsunoda T, Tanaka M, Oikawa M (1975) Degradation of polyacrylamide molecules in
965 aqueous-solutions by high-speed stirring. Bulletin of the Chemical Society of Japan 48:2597-
966 2598

967 Nakamiya K, Kinoshita S (1995) Isolation of polyacrylamide-degrading bacteria. Journal of
968 Fermentation and Bioengineering 80:418-420

969 Nawaz MS, Franklin W, Cerniglia CE (1993) Degradation of acrylamide by immobilized cells of a
970 Pseudomonas sp. and Xanthomonas maltophilia. Can. J. Microbiol. 39:207-212

971 Nawaz MS, Khan AA, Seng JE, Leakey JE, Siitonen PH, Cerniglia CE (1994) Purification and
972 characterization of an amidase from an acrylamide-degrading Rhodococcus sp. Appl Environ
973 Microbiol 60:3343-8

974 Nawaz MS, Khan AA, Bhattacharayya D, Siitonen PH, Cerniglia CE (1996) Physical, biochemical, and
975 immunological characterization of a thermostable amidase from Klebsiella pneumoniae NCTR
976 1. J. Bacteriol. 178:2397-2401

977 Nawaz MS, Billedeau SM, Cerniglia CE (199): Influence of selected physical parameters on the
978 biodegradation of acrylamide by immobilized cells of Rhodococcus sp. Biodegradation 9:381-
979 387

980 Nguyen QD, Boger DV (1985) Thixotropic behaviour of concentrated bauxite residue suspensions
981 Rheol Acta 24:427-437

982 Nguyen QD, Boger DV (1998) Application of rheology to solving tailings disposal problems.
983 International Journal of Mineral Processing 54:217-233

984 Ohtaki A, Murata K, Sato Y, Noguchi K, Miyatake H, Dohmae N, Yamada K, Yohda M, Odaka M
985 (2010) Structure and characterization of amidase from *Rhodococcus* sp. N-771: Insight into
986 the molecular mechanism of substrate recognition. *Biochim. Biophys. Acta* 1804:184-192

987 Parfitt RL, Greenland JD (1970) Adsorption of polysaccharides by montmorillonite. *Soil Science*
988 *Society of America Journal* 34:562-866

989 Paterson RW, Abernathy FH (1970) Turbulent flow drag reduction and degradation with dilute polymer
990 solutions. *J. Fluid. Mech* 43:689-710

991 Pefferkorn E, Nabzar L, Varoqui R (1987) Polyacrylamide Na-kaolinite interactions – effect of
992 electrolyte concentration on polymer adsorption. *Colloid and Polymer Science* 265:889-896

993 Pereira AS, Soares EJ (2012) Polymer degradation of dilute solutions in turbulent drag reducing flows
994 in a cylindrical double gap rheometer device. *J. Non Newtonian Fluid Mech.* 179-180:9-22

995 Potts JR, Clarke PH (1976) The effect of nitrogen limitation on catabolite repression of amidase,
996 histidase and urocanase in *Pseudomonas aeruginosa*. *J Gen Microbiol* 93:377-87

997 Povkh LI, Chernyuk VV (1986) An experimental investigation of the effect of a polyacrylamide additive
998 on the pressure drop in an expansion *Journal of engineering physics* 51:1001-1004

999 Pradip YAA, Fuerstenau DW (1980) The adsorption of polyacrylamide flocculants on apatites. *Colloid*
1000 *and Polymer Science* 258:1343-1353

1001 Rabek JF (1996) In *Photodegradation of Polymers: Physical Characteristics and Applications*.
1002 Springer-Verlag. Berlin

1003 Rasteiro MG, Garcia FAP, Ferreira P, Blanco A, Negro C, Antunes E (2008) Evaluation of flocs
1004 resistance and reflocculation capacity using the LDS technique. *Powder Technology* 183:231-
1005 238

1006 Ren G-m, Sun D-z, Chung Jong S (2006) Kinetics study on photochemical oxidation of polyacrylamide
1007 by ozone combined with hydrogen peroxide and ultraviolet radiation. *Journal of Environmental*
1008 *Sciences-China* 18:660-664

1009 Rho T, Park J, Kim C, Yoon HK, Suh HS (1996) Degradation of polyacrylamide in dilute solution.
1010 *Polymer Degradation and Stability* 51:287-293

1011 Ruehrwein RA, Ward DW (1952) Mechanism of clay aggregation by polyelectrolytes. *Soil Science*
1012 73:485-492

1013 Santoshkumar M, Nayak AS, Anjaneya O, Karegoudar TB (2010) A plate method for screening of
1014 bacteria capable of degrading aliphatic nitriles. *J. Ind. Microbiol. Biotechnol.* 37:111-115

1015 Sathesh Prabu C, Thatheyus AJ (2007) Biodegradation of acrylamide employing free and immobilized
1016 cells of *Pseudomonas aeruginosa*. *International Biodeterioration & Biodegradation* 60:69-73

1017 Sato T, Ruch R (1980) Stabilization of colloidal dispersions by polymer adsorption. In: Dekker (Editor),
1018 Surfactant science serie. Marcel Dekker Inc., New York

1019 Scott JP, Fawell PD, Ralph DE, Farrow JB (1996) The shear degradation of high-molecular-weight
1020 flocculant solutions. *Journal of Applied Polymer Science* 62:2097-2016

1021 Senden TJ, Di Meglio JM, Silberzan I (2000) The conformation of adsorbed polyacrylamide and
1022 derived polymers. *Comptes Rendus De L Academie Des Sciences Serie Iv Physique*
1023 *Astrophysique* 1 :1143-1152

1024 Seybold CA (1994) Polyacrylamide review: Soil conditioning and environmental fate. *Communications*
1025 *in Soil Science and Plant Analysis* 25:2171-2185

1026 Shanker R, Ramakrishna C, Seth PK (1990) Microbial degradation of acrylamide monomer. *Arch.*
1027 *Microbiol.* 154:192-198

1028 Sharma M, Sharma N, Bhalla T (2009) Amidases: versatile enzymes in nature. *Reviews in*
1029 *Environmental Science and Biotechnology* 8:343-366

1030 Shukor MY; Gusmanizar N; Azmi NA et al. (2009a) Isolation and characterization of an acrylamide-
1031 degrading *Bacillus cereus*. *Journal of environmental biology.* 30(1)57-64

1032 Shukor MY, Gusmanizar N, Ramli J, Shamaan NA, MacCormack WP, Syed MA (2009b) Isolation and
1033 characterization of an acrylamide-degrading Antarctic bacterium. *J. Environ. Biol.* 30:107-112

1034 Smith, EA and Oehme FW (1993) Rapid direct analysis of acrylamide residue in polyacrylamide
1035 thickening agents by HPLC. *Journal of Chromatographic Science* 31:192-195.

1036 Smith EA, Prues SL, Oehme FW (1996) Environmental degradation of polyacrylamides .1. Effects of
1037 artificial environmental conditions: Temperature, light, and pH. *Ecotoxicology and*
1038 *Environmental Safety* 35:121-135

1039 Smith EA, Prues SL, Oehme FW (1997) Environmental degradation of polyacrylamides .2. Effects of
1040 environmental (outdoor) exposure. *Ecotoxicology and Environmental Safety* 37:76-91

1041 Sojka RE, Entry JA (2000) Influence of polyacrylamide application to soil on movement of
1042 microorganisms in runoff water. *Environ Pollut* 108:405-12

1043 Sojka RE, Bjorneberg DL, Entry JA, Lentz RD, Orts WJ (2007) Polyacrylamide in Agriculture and
1044 Environmental Land Management. *Advances in Agronomy* 92:75-162

1045 Spicer PT, Pratsinis SE, Raper J, Amal R, Bushell G, Meesters G (1998) Effect of shear schedule on
1046 particle size, density, and structure during flocculation in stirred tanks. *Powder Technology*
1047 97:26-34

1048 Spofford TL, Pfeiffer KL (1996) Agricultural irrigation polyacrylamide application standard, *Managing*
1049 *Irrigation-Induced Erosion and Infiltration with Polyacrylamide*, College of Southern Idaho,
1050 Twin Falls, Id USA

1051 Stephens SH (1991) Final report on the safety of polyacrylamide. *Journal of the American college of*
1052 *toxicology* 10:193-202

1053 Stutzmann T, Siffert B (1977) Contribution to adsorption mechanism of acetamide and polyacrylamide
1054 on to clays. *Clays and Clay Minerals* 25:392-406

1055 Suzuki K, Sugiyama A, Nakazato K (1978) Study of acrylamide-gel chromatography 3 spherical gels of
1056 polyacrylamide imdized by heat-treatment. *Nippon Kagaku Kaishi*, 1385-1389

1057 Suzuki K, Nakazato K, Takasaki T (1979) Study of acrylamide-gel chromatography 4 spherical gels of
1058 polyacrylamide co-polymerized with acrylonitrile. *Nippon Kagaku Kaishi*, 1327-1331

1059 Syed MA, Ahmad SA, Kusnin N, Shukor MY (2012) Purification and characterization of amidase from
1060 acrylamide-degrading bacterium *Burkholderia* sp. strain DR.Y27. *African Journal of*
1061 *Biotechnology* 11:329-336

1062 Tanaka H, Odberg L, Wagberg L, Lindstrom T (1990) Adsorption of cationic polyacrylamide onto
1063 monodisperses polystyrene lattices and cellulose fiber – effect of molecular-weight and
1064 charge-density of cationic polyacrylamides. *Journal of Colloid and Interface Science* 13: 219-
1065 228

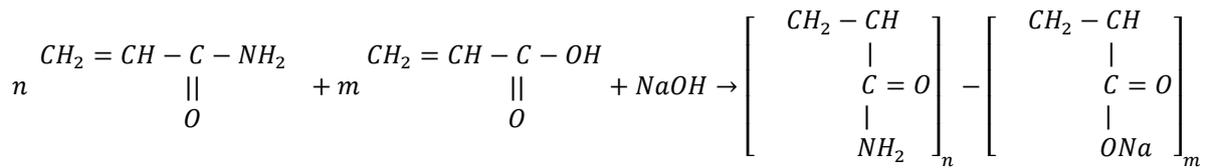
1066 Tang S, Ma Y, Shiu C (2001) Modeling the mechanical strength of fractal aggregates. *Colloids Surf.*
1067 *Physicochem. Eng. Aspects* 180:7-16

1068 Taylor ML, Morris GE, Self PG, Smart RS (2002) Kinetics of adsorption of high molecular weight
1069 anionic polyacrylamide onto kaolinite: The flocculation process. *Journal of Colloid and*
1070 *Interface Science* 250: 28-36

- 1071 Tekin N, Demirbas O, Alkan M (2005) Adsorption of cationic polyacrylamide onto kaolinite.
1072 Microporous and Mesoporous Materials 85:340-350
- 1073 Tekin N, Dincer A, Demirbas O, Alkan M (2006) Adsorption of cationic polyacrylamide onto sepiolite.
1074 Journal of Hazardous Materials 134:211-219
- 1075 Tekin N, Dincer A, Demirbas O, Alkan M (2010) Adsorption of cationic polyacrylamide (C-PAM) on
1076 expanded perlite. Applied Clay Science 50:125-129
- 1077 Thanyacharoen U, Tani A, Charoenpanich J (2012) Isolation and characterization of Kluyvera
1078 georgiana strain with the potential for acrylamide biodegradation. J Environ Sci Health A Tox
1079 Hazard Subst Environ Eng 47:1491-9
- 1080 Theng BKG (1979) Formation and properties of clay-polymer complexes. Developement in Soil
1081 Sciences 9:9-11
- 1082 Theng BKG (1982) Clay-polymer interactions - summary and perspectives. Clays and Clay Minerals
1083 30:1-10
- 1084 Togola A, Coureau C, Guezennec AG, Touzé S (Submitted 2014). A sensitive analytical procedure for
1085 acrylamide in environmental water samples by offline SPE-UPLC/MS/MS. Environ Sci Pollut
1086 Res (SI AQUAPOL)
- 1087 Touzé S, Guerin V, Guezennec AG, Binet S, Togola A (2014). Dissemination of acrylamide monomer
1088 from polyacrylamide based flocculant use - sand and gravel quarry case study. Environ Sci
1089 Pollut Res (SI AQUAPOL)
- 1090 UNPG (2013) Polyacrylamide based flocculant used in quarries (in French). Union National des
1091 Producteurs de Granulats UNPG
- 1092 Van de Ven TGM (1994) Kinetic aspects of polymer and polyelectrolyte adsorption on surfaces.
1093 Advances in Colloid and Interface Science 48:121-140
- 1094 Ver Vers LM (1999) Determination of acrylamide monomer in polyacrylamide degradation studies by
1095 high-performance liquid chromatography. Journal of Chromatographic Science 37:486-494
- 1096 Vlachogiannis M, Liberatore MW, McHugh AJ, Hanratty TJ (2003) Effectiveness of a drag reducing
1097 polymer: Relation to molecular weight distribution and structuring. Physics of Fluids 15:3786-
1098 3794
- 1099 Wade JHT, Kumar P (1972) Electronic microscope studies of polymer degradation. J. Hydronaut. 6-40

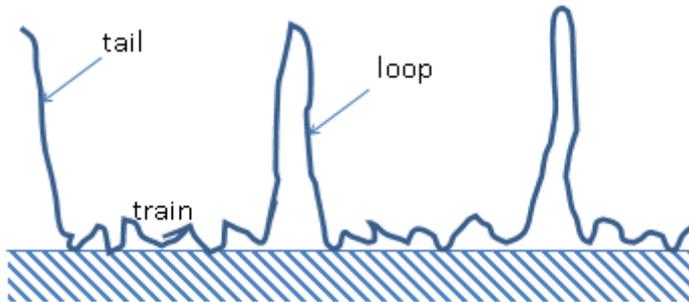
- 1100 Wakaizumi M, Yamamoto H, Fujimoto N, Ozeki K (2009) Acrylamide degradation by filamentous fungi
1101 used in food and beverage industries. *J. Biosci. Bioeng.* 108:391-3
- 1102 Wampler DA, Ensign SA (2005) Photoheterotrophic metabolism of acrylamide by a newly isolated
1103 strain of *Rhodospseudomonas palustris*. *Appl. Environ. Microbiol.* 7: 5850-5857
- 1104 Weideborg M, Kallqvist T, Odegard KE, Sverdrup LE, Vik EA (2001) Environmental risk assessment of
1105 acrylamide and methylolacrylamide from a grouting agent used in the tunnel construction of
1106 Romeriksporten, Norway. *Water Research* 35:2645-2652
- 1107 Wen Q, Chen Z, Zhao Y, Zhang H, Feng Y (2010) Biodegradation of polyacrylamide by bacteria
1108 isolated from activated sludge and oil-contaminated soil. *J. Hazard. Mater.* 175:955-959
- 1109 Wilson S, Drew R (1991) Cloning and DNA sequence of *amiC*, a new gene regulating expression of
1110 the *Pseudomonas aeruginosa* aliphatic amidase, and purification of the *amiC* product. *J*
1111 *Bacteriol* 173:4914-21
- 1112 Wilson SA, Wachira SJ, Drew RE, Jones D, Pearl LH (1993) Antitermination of amidase expression in
1113 *Pseudomonas aeruginosa* is controlled by a novel cytoplasmic amide-binding protein. *Embo J*
1114 12:3637-42
- 1115 Wilson SA, Drew RE (1995) Transcriptional analysis of the amidase operon from *Pseudomonas*
1116 *aeruginosa*. *J Bacteriol* 177:3052-7
- 1117 World Health Organization (1985) Environmental Health Criteria 1985, 49, 1. International programme
1118 on chemical safety Web. <http://www.inchem.org/documents/ehc/ehc/ehc49.htm>
- 1119 Woodrow JE, Seiber JN, Miller GC (2008) Acrylamide release resulting from sunlight irradiation of
1120 aqueous polyacrylamide/iron mixtures. *Journal of Agricultural and Food Chemistry* 56:2773-
1121 2779
- 1122 Yamada H, Asano Y, Hino T, Tani Y (1979) Microbial utilization of acrylonitrile. *Journal of fermentation*
1123 *technology* 5: 8-14
- 1124 Yeung A, Gibbs A, Pelton R (1997) Effect of shear on the strength of polymer-induced flocs. *Journal of*
1125 *Colloid and Interface Science* 196:113-115
- 1126 Yeung AKC, Pelton R (1996) Micromechanics: A new approach to studying the strength and breakup
1127 of flocs. *Journal of Colloid and Interface Science* 184:579-585

1128 Young MH, Tappen JJ, Miller GC, Carroll S., Susfalk R.B. (2007). Risk Characterization: Using Linear
1129 Anionic Polyacrylamide to Reduce Water Seepage from Unlined Water Delivery Canal
1130 Systems. Publication no. 41226. Desert Research Institute, Las Vegas, NV.
1131



1132 **Fig. 1 Synthesis pathway of anionic PAMs**

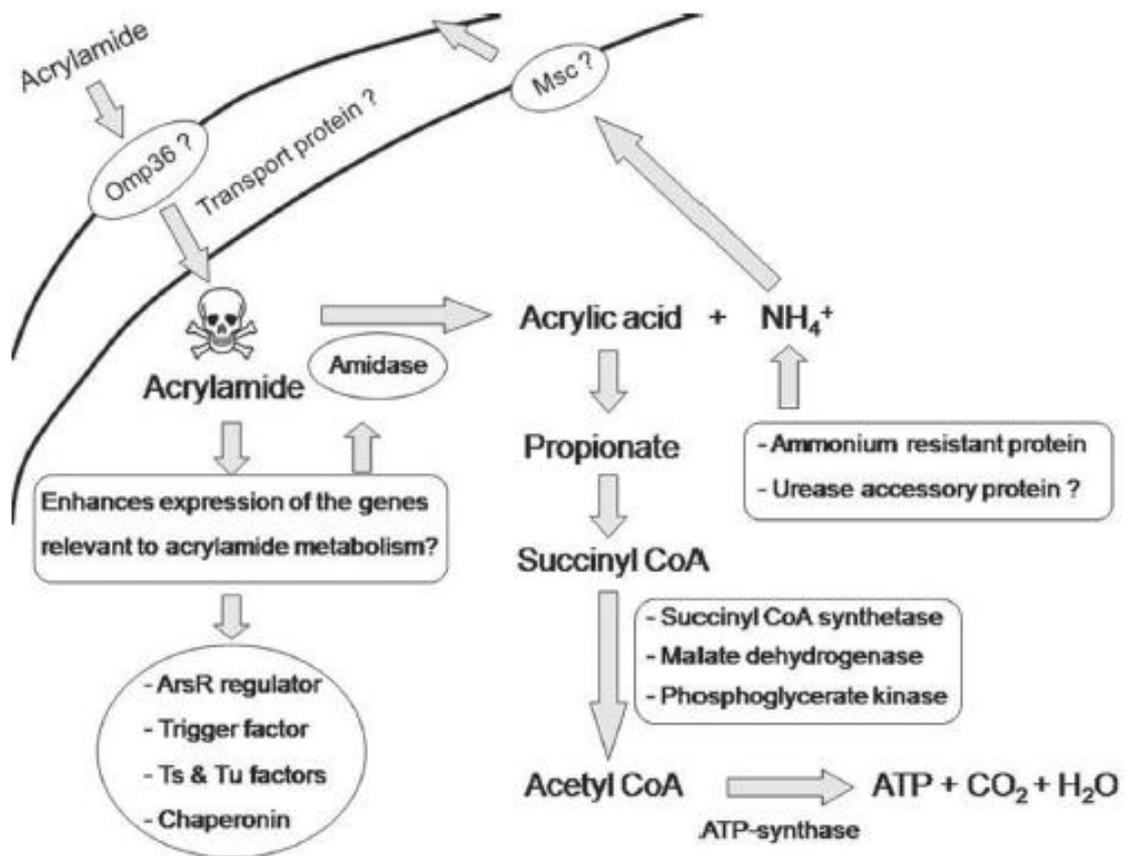
1133



1134

1135 **Fig. 2 Conformation of a polymer adsorbed onto the surface of a mineral particle**

1136



1137

1138 **Fig. 3 Metabolic pathway of AMD degradation in *Enterobacter aerogenes* proposed by**
 1139 **Charoenpanich and Tani (2014).**

1140

1141 **Table 1 Aerobic biodegradation of acrylamide in various environmental samples**

References	Acrylamide concentration	Temperature	Lag time	Time for complete AMD degradation
Labahn <i>et al.</i> (2010)	50 µg.L ⁻¹	15°C	No degradation observed after 72 h	
		25°C	0-12h	24 h
Croll (1973)	10 µg.L ⁻¹	Non indicated	5h-50h	50h-300h
Brown (1979)	5 mg.L ⁻¹	20°C	75h	125 h
Vers (1999)	1,5-2,5 mg.L ⁻¹	Non indicated	0h	from 480 h to more than 1100 h

1142

1143

1144 **Table 2 Molecular weight, pH and temperature optima of amidases purified from AMD-**
 1145 **degrading bacteria.**

Bacterial strains	Molecular weight (kDa)	Optimum pH	Optimum temperature	Reference
<i>Geobacillus thermoglucosidasius</i> AUT-01	38.0	6.2	70°C	Cha and Chambliss, 2013
<i>Geobacillus pallidus</i> BTP-5x MTCC 9225	38.5	8.0	60°C	Sharma <i>et al.</i> 2012
<i>Ralstonia eutropha</i> AUM-01	38.0	6.3	55°C	Cha and Chambiss, 2011
<i>Variovorax boronicumulans</i> CGMCC 4969	42.0	ND*	ND*	Liu <i>et al.</i> 2013
<i>Burkholderia</i> sp. DR.Y27	47.0	8.0	40°C	Syed <i>et al.</i> 2012
<i>Rhodococcus</i> sp.	44.5	8.5	40°C	Nawaz <i>et al.</i> 1994
<i>Rhodococcus</i> sp. N-771	54.7	5.9	55°C	Ohtaki <i>et al.</i> 2010
<i>Klebsiella pneumoniae</i> NCTR 1	62.0	7.0	65°C	Nawaz <i>et al.</i> 1996
<i>Geobacillus pallidus</i> RAPc8	38.0	7.0	50°C	Makhongela <i>et al.</i> 2007
<i>Pseudonocardia thermophila</i> DSMZ 43832	52.0	7.0	70°C	Egorova <i>et al.</i> 2004
<i>Rhodococcus erythropolis</i> TA37	55.0	8.0	55°C	Lavrov <i>et al.</i> 2010

1146 * ND: Not Determined

1147

1148